

Filed on behalf of Junior Party

Paper No. \_\_\_\_

THE REGENTS OF THE UNIVERSITY OF CALIFORNIA,  
UNIVERSITY OF VIENNA, AND EMMANUELLE CHARPENTIER

By: Eldora L. Ellison, Ph.D., Esq.  
Eric K. Steffe, Esq.  
David H. Holman, Ph.D., Esq.  
Byron L. Pickard, Esq.  
Paul A. Ainsworth, Esq.  
John Christopher Rozendaal, Esq.  
Michael E. Joffre, Ph.D., Esq.  
Pauline M. Pelletier, Esq.  
STERNE, KESSLER, GOLDSTEIN & FOX P.L.L.C.  
1100 New York Avenue, NW  
Washington, D.C. 20005  
Tel: (202) 371-2600  
Fax: (202) 371-2540  
eellison-PTAB@sternekessler.com  
esteffe-PTAB@sternekessler.com  
dholman-PTAB@sternekessler.com  
bpickard-PTAB@sternekessler.com  
painsworth-PTAB@sternekessler.com  
jcrozendaal-PTAB@sternekessler.com  
mjoffre-PTAB@sternekessler.com  
ppelletier-PTAB@sternekessler.com

By: Li-Hsien Rin-Laures, M.D., Esq.  
RINLAURES LLC  
321 N. Clark Street, 5th floor  
Chicago, IL 60654  
Tel. (773) 387-3200  
Fax (773) 929-2391  
lily@rinlauresip.com

Sandip H. Patel, Esq.  
MARSHALL GERSTEIN & BORUN LLP  
6300 Willis Tower,  
233 South Wacker Drive  
Chicago, IL 60606  
Tel: (312) 474-6300  
Fax: (312) 474-0448  
spatel@marshallip.com

UNITED STATES PATENT AND TRADEMARK OFFICE

BEFORE THE PATENT TRIAL AND APPEAL BOARD

**THE REGENTS OF THE UNIVERSITY OF CALIFORNIA, UNIVERSITY  
OF VIENNA, AND EMMANUELLE CHARPENTIER**

Applications 15/947,680; 15/947,700; 15/947,718; 15/981,807; 15/981,808;  
15/981,809; 16/136,159; 16/136,165; 16/136,168; 16/136,175; 16/276,361;  
16/276,365; 16/276,368; and 16/276,374,  
Junior Party,

v.

**SIGMA-ALDRICH, CO., LLC**  
Application 15/456,204

Senior Party.

Patent Interference No. 106,132 (DK)  
(Technology Center 1600)

**CVC REPLY MOTION 3  
(to change the Count)**

**TABLE OF CONTENTS**

I. The Motion Will Not be Moot if the PTAB Grants Sigma’s Substantive Motion 1 ..... 1

II. The PTAB Should Exercise its Discretion to Change the Scope of the Count to Better Reflect the Counts in the Related Interferences. .... 2

III. Sigma Fails to Establish That its Best Proofs Fall Outside the Scope of the Count. .... 4

IV. Substituting the Count Will Streamline the Issues in the Interference. .... 5

V. Granting CVC’s Motion 3 Will Not Impact the Designation of Claims, Priority Benefit, and Patentability Over the Art. .... 7

VI. Conclusion ..... 7

**TABLE OF AUTHORITIES**

**Cases**

*Hitzeman v. Rutter*,  
243 F.3d 1345 (Fed. Cir. 2001).....2

*Hunt v. Treppschuh*,  
523 F.2d 1386 (C.C.P.A. 1975) .....4

*In re Vivint, Inc.*,  
14 F.4th 1342 (Fed. Cir. 2021) .....2, 3

*Reagents of the University of California v. The Broad Institute*,  
Interference No. 106,115 (P.T.A.B. Sept. 10, 2020). .....1

**Other Authorities**

37 C.F.R. § 41.1(b) .....3

37 C.F.R. § 41.203(b) .....2

37 C.F.R. § 41.125(a).....2

1    **INTRODUCTION**

2           The count in this interference should reflect the common invention defining multiple  
3 pending, related interferences - single-guide CRISPR-Cas9 systems useful in eukaryotic cells.  
4    *See* SO ¶ 208.2. CVC’s half of Count 1 aligns with the counts in the interferences between CVC  
5 and Broad (Interference No. 106,115), Ex. 2560, 12-13; CVC and ToolGen (Interference No.  
6 106,127), Ex. 2561 at 5-6; and Broad and ToolGen (Interference No. 106,126), Ex. 2562, 12-13;  
7 MF 1. CVC’s halves of the counts and CVC’s involved claims in those interferences are the  
8 same as in this one. Broad’s and ToolGen’s halves of the counts in those interferences are also  
9 commensurate in scope with CVC’s half of the count here. However, that is not the case in this  
10 interference - Sigma’s half of Count 1 is not aligned with CVC’s half or any other parties’  
11 halves. Sigma argues that this misalignment is justified because it has generic-guide claims,  
12 ignoring that CVC also has such generic-guide claims involved in this interference. In the ’115  
13 interference, the PTAB specifically rejected the argument that alleging dual-guide proofs  
14 mandates a generic-guide count. *See Regents of the University of California v. The Broad*  
15 *Institute*, Int. No. 106,115 Paper 877 (Decision on Motions) at 39:22-40:8 (P.T.A.B. Sept. 10,  
16 2020). There is no reason for the PTAB to treat Sigma—and Sigma alone—differently than  
17 CVC, Broad, and ToolGen. The PTAB should grant CVC Motion 3 and substitute a count  
18 aligned with the other interferences.

19    ***I. The Motion Will Not be Moot if the PTAB Grants Sigma’s Substantive Motion 1***

20           On page 3 line 5 through page 4 line 13, Sigma argues that CVC’s substantive Motion 3  
21 (to change the count) would be moot if the Board grants Sigma’s Substantive Motion 1 (to  
22 change the count). The reply is that CVC’s Motion 3 will not be moot because whether or not the  
23 scope of the interference is limited to cleavage *plus* donor integration (as Sigma requests in its  
24 Motion 1) is a separate issue that does not impact whether or not the Count should recite single-

1 guide RNA. Sigma's Proposed Count 2 propagates the mismatched guide RNA format: CVC's  
2 half of the count is directed to *single-guide* RNA, while Sigma's half of its proposed count is  
3 directed to *generic guide* RNA (encompassing both dual-molecule and single-molecule RNAs).  
4 CVC's Motion 3 proposes to change Sigma's half of the count so that it too recites a single-  
5 molecule RNA, making the CVC and Sigma halves of the count consistent.

6 Accordingly, the relief requested in CVC's Motion 3 would not be mooted by the grant of  
7 Sigma's Motion 1 as Sigma argues. It would, however, be mooted by any disposition of Sigma's  
8 Motion 1 that results in a judgment in favor of CVC, as explained in CVC Opposition 1. If the  
9 PTAB does not enter such judgment at this time, then it should grant CVC's Motion 3 to  
10 substitute the count *regardless* of how it decides Sigma's Motion 1.

11 ***II. The PTAB Should Exercise its Discretion to Change the Scope of the Count to Better***  
12 ***Reflect the Counts in the Related Interferences.***

13 The PTAB has discretion to substitute the count in an interference. *See Hitzeman v.*  
14 *Rutter*, 243 F.3d 1345, 1359 (Fed. Cir. 2001) (finding no abuse of discretion in narrowing the  
15 scope of a count). Sigma identifies no contrary authority. Sigma cites no authority refuting that  
16 general principles of consistency, fairness, and clarity justify exercising that discretion—  
17 especially where the Federal Circuit has so recently endorsed following these general principles  
18 in administering Patent Office proceedings. *See In re Vivint, Inc.*, 14 F.4th 1342, 1351 (Fed. Cir.  
19 2021) (supporting the general proposition that the PTAB should not act arbitrarily and  
20 capriciously across multiple proceedings).

21 On page 6 line 11 through page 10 line 4, Sigma argues that the count here should differ  
22 from the counts in the related interferences because Sigma is the only party to allege proofs  
23 relating to both single-guide and dual-molecule RNA. Sigma is wrong. Broad also alleged such  
24 proofs (which CVC disputed), but the PTAB did not find Broad's allegations a sufficiently

1 compelling reason to broaden the '115 interference count beyond single-guide RNA. *See* Ex.  
2 2400, 36:6-8. Broad even alleged that its dual-molecule proofs were earlier than its single-guide  
3 proofs, *id.*, something Sigma does not even allege. CVC also has claims directed to both single-  
4 guide and dual-molecule RNAs in all pending interferences, yet its half of the count has always  
5 been limited to single-guide. Ex. 2560, 14-16; Ex. 2561, 7-10; *see also* Ex. 2655, 109:16-117:19  
6 (admitting that CVC has involved dual-guide claims and not disputing that CVC has involved  
7 generic-guide claims). Sigma's entire rebuttal to CVC's argument for consistency and fairness  
8 across proceedings is thus based on the false premise that it is uniquely situated.

9         The PTAB should treat similarly situated parties similarly across multiple related  
10 proceedings. *See Vivint*, 14 F.4th at 1351. Such consistency is only achieved here by granting  
11 CVC's motion, which presents a count where Sigma's half (Sigma claim 33) recites limitations  
12 that match the single-guide aspect of CVC's half of the count in this interference as well as  
13 CVC's, Broad's, and ToolGen's halves of the counts of similar related interferences. 37 C.F.R. §  
14 41.1(b). Administering the interference with CVC's proposed count will provide the public a  
15 clearer record as to who was first to invent the common subject matter involved in these related  
16 interferences among the multiple parties involved.

17         Substituting CVC's proposed count will also limit the inherent prejudice to CVC that  
18 results from arbitrarily giving Sigma special treatment. Sigma requested this interference, which  
19 the PTAB declared long after having declared other interferences involving all of the same CVC  
20 applications and claims. CVC is litigating materially the same count for all of its involved claims  
21 across those other interferences. Yet the count in this interference (not limited to sgRNA) is the  
22 only one different from the other interferences (limited to sgRNA). Any arbitrary and undeserved  
23 treatment afforded to Sigma in this interference necessarily prejudices CVC. CVC has  
24 sufficiently shown why all of these interferences should be administered consistently. Sigma

1 offered no compelling reason or authority to administer this interference differently. The PTAB  
2 should therefore substitute Sigma's claim 31 with claim 33 of Sigma's '204 Application in the  
3 count.

4 ***III. Sigma Fails to Establish That its Best Proofs Fall Outside the Scope of the Count.***

5 For all its accusations that CVC is attempting to hamstring Sigma by excluding Sigma's  
6 best proofs, *see, e.g.*, Paper 708, Sigma Opp. at 5:8-10, Sigma failed to make a proffer that its  
7 best proofs fall outside the scope of CVC's proposed count. To the contrary, Sigma agrees that  
8 the same gel evidencing its alleged invention includes lanes for both single-guide and dual-  
9 molecule RNAs. Indeed, Sigma's counterarguments justifying its "outlier" treatment rely on  
10 "both" its alleged single-guide and dual-guide proofs equally. *Id.* at 11:13-12:17. Because it need  
11 only show one embodiment within the scope of the count to prevail on priority, Sigma cannot be  
12 prejudiced by narrowing the count to single-guide RNA unless doing so would *exclude* its best  
13 proofs. *See Hunt v. Treppschuh*, 523 F.2d 1386, 1389 (C.C.P.A. 1975). Sigma's repeated  
14 assertion that its proofs include *both* single-guide and dual-guide RNA fails to establish that its  
15 *best* proofs are confined to dual-guide RNA. Paper 708 at 11:13-12:17.

16 Sigma does not allege that its single-guide proofs relying on FACS experiments may be  
17 inadequate. Sigma's only purported difference between its single-guide and dual-molecule  
18 proofs is that it has later, confirmatory PCR evidence regarding the dual-molecule experiments  
19 that it does not have for the single-guide experiments. Paper 708, Sigma Opp at 7:11-24; MF 63.  
20 Sigma offers no evidence that PCR evidence is necessary. Indeed, Sigma's own expert dismisses  
21 Sigma's PCR data as merely "further confirmation" and does not rely on it. Ex.1080, ¶ 79 n.1.  
22 Sigma also does not allege that its alleged date of invention would be earlier or better-  
23 corroborated for its dual-guide proofs than its single-guide proofs. MF 64. Sigma thus has not

1 established that it will suffer any prejudice if the Board substitutes CVC’s Proposed Court 2 for  
2 current Count 1.

3 ***IV. Substituting the Count Will Streamline the Issues in the Interference.***

4 Sigma’s involved application is ambiguous as to the scope of the “guide RNA” recited in  
5 Sigma’s claim 31, a claim construction the PTAB may justifiably avoid by defining Sigma’s half  
6 of the count as claim 33. Specifically, Sigma’s Claim 31 recites “guide RNA” without specifying  
7 whether the guide RNA is a single molecule (with three regions) as described in the  
8 specification. Claim 33 depends from claim 31 and specifies that the guide RNA is a single  
9 molecule. Changing the scope of the count to recite Sigma’s claim 33 instead of Sigma’s claim  
10 31 will clarify that the common, interfering subject matter is a sgRNA CRISPR-Cas9 system in a  
11 eukaryotic cell. It will also simplify the interference by focusing on who was the first to invent  
12 sgRNA CRISPR-Cas9 in a eukaryotic cell, and eliminate any dispute over whether Sigma’s  
13 alleged dual-guide proofs actually satisfy Sigma’s own definition of “guiding RNA” and fall  
14 within the scope of a count that recites “guide RNA.”

15 Sigma’s argument that the term “guide RNA” used in claim 31 includes both single-guide  
16 and dual-guide RNAs is based on a description of two-molecule “guiding RNA.” Paper 708,  
17 Sigma Opp at 12-13 (citing Sigma’s P1 at ¶¶ [0019], [0023]). Specifically, Sigma’s involved  
18 application defines “the guiding RNA” as comprising a first single-stranded region, “a second  
19 internal region that forms a stem loop structure,” and a third single-stranded region. Ex. 2026,  
20 ¶¶ [0021], [0019]; MF 9; Cf. Ex. 2031, Fig. 5A (annotated) (cited in Sigma’s ’204 application);  
21 Ex. 2549, ¶ 99; Ex. 2585, ¶¶ [0071]-[0072]. This structure as described is necessarily a single-  
22 guide RNA because the crRNA and tracrRNA are physically linked by loop to form a “stem loop  
23 structure.” MF 9. When so linked, they must constitute a single molecule. MF 65.



1           At best, Sigma points to *some* intrinsic evidence that is confusing in view of the  
2 definition of guide RNA in the application. Sigma points to ¶ [0023] of its specification, which  
3 notes that each “guiding RNA” can alternatively comprise two molecules, each of which  
4 includes a portion of the stem from the previously described stem loop structure. Paper 708,  
5 Sigma Opp at 13:22-28. This paragraph, however, conflicts with the overall weight of the  
6 intrinsic evidence regarding the scope of the term “guide RNA,” and therefore does not establish  
7 that a POSA would construe the term “guide RNA” more broadly.

8           Sigma argues, falsely, that “CVC and its expert (Dr. Scott Bailey) fail to consider the  
9 [language of] subsequent ¶ [0023], which unambiguously explains that the guide RNA—  
10 including a ‘stem loop’—can be a dual-molecule guide RNA.” *Id.* at 13:20-22. Therein lies  
11 Sigma’s problem: There is no physically possible guide-RNA structure that is both (a) a dual-  
12 molecule guide RNA and (b) a guide RNA with “a second internal region that forms a stem loop  
13 structure”, i.e., comprising a linker loop as defined by Sigma’s specification. *See Ex. 1082, 85:4-*  
14 *86:6* (explaining that the only way a guide RNA can comprise two molecules is if it does *not*  
15 have an intact stem loop in the second region); MF 66.

16           Sigma’s application—as Sigma urges the PTAB to read it here—thus contradicts itself.  
17 Sigma’s expert did not consider that contradiction. *See Ex.2655, 119:16-127:13* (admitting that  
18 she offered no analysis of the structure of the single-guide or dual-guide RNA, and refusing to  
19 provide that analysis during her cross-examination); MF 67. CVC’s expert Bailey did consider  
20 the contradictory language. According to Bailey, a POSA would resolve the inconsistency in  
21 Sigma’s specification by reading the three-region single-molecule definition of “guiding RNA”  
22 to describe the claimed “guide RNA” and treating the inadequately explained dual-molecule as  
23 another way of describing the single-guide structure (which does “comprise two separate  
24 molecules” that are then joined with a linker loop), or an unclaimed alternative. *See Ex 2549, ¶¶*

1 98, 102 (explaining how a POSA would interpret Sigma’s “ambiguous” and “inconsistent”  
2 descriptions of the guiding RNA).

3 The intrinsic evidence Sigma cites only supports construing the guide RNA recited in  
4 claim 31 as single-guide RNA. Substituting the count to expressly recite that which Sigma is  
5 properly limited to through claim construction anyway is yet another reason why Sigma is not  
6 prejudiced by this Motion. The economy of avoiding this dispute and focusing the issues for the  
7 priority phase on the common inventive subject matter further supports ensuring Sigma’s half of  
8 the count is Sigma claim 33.

9 ***V. Granting CVC’s Motion 3 Will Not Impact the Designation of Claims, Priority Benefit,***  
10 ***and Patentability Over the Art.***

11 Ordinarily, substitution of a count would include an assessment of (a) whether the count  
12 is patentable over prior art, (b) which of each party’s claims correspond to the count, and (c)  
13 each party’s accorded benefit. But, the substitution of CVC’s proposed count has no impact on  
14 any of these analyses. Sigma does not dispute that the substitution of the count has no impact.  
15 Sigma also does not dispute that the analysis and relief requested in CVC Substantive Motion 4  
16 to Add the Claims of Sigma Patent No. 10,731,181 (“the ’181 patent) and No. 10,745,716 (“the  
17 ’716 patent”) to this Interference are equally applicable if Count 1 is substituted with CVC’s  
18 proposed count. These additional considerations all weigh in favor of granting CVC’s Motion 3.

19 ***VI. Conclusion***

20 The PTAB should grant CVC’s Motion 3 or ensure that whatever count it may substitute  
21 is limited to a sgRNA CRISPR-Cas 9 system, consistent with multiple, related, pending  
22 interferences.

23

1

Respectfully submitted,

By /Eldora L. Ellison/  
Eldora L. Ellison, Ph.D., Esq.  
Lead Attorney for UC and UV  
Registration No. 39,967  
STERNE, KESSLER, GOLDSTEIN & FOX P.L.L.C.  
1100 New York Avenue, NW  
Washington, D.C. 20005

By /Li-Hsien Rin-Laures/  
Li-Hsien Rin-Laures, M.D., Esq.  
Lead Attorney for EC  
Registration No. 33,547  
RINLAURES LLC  
321 N. Clark Street, 5th floor  
Chicago, IL 60654

Date: April 7, 2022

Date: April 7, 2022

2

**APPENDIX A: EXHIBIT LIST**

Exhibit No.	Description
1080	Cannon Supp'l Decl.
1082	Bailey Tr. 01/24/22
2006	U.S. Appl. No. 14/685,504, filed April 13, 2015
2007	U.S. Appl. No. 15/138,604, filed April 16, 2016
2009	Prov. Appl. No. 61/652,086, filed May 25, 2012
2010	Prov. Appl. No. 61/716,256, filed October 19, 2012
2011	Prov. Appl. No. 61/757,640, filed January 28, 2013
2015	U.S. Appl. No. 13/842,859, filed March 15, 2013
2017	U.S. Patent No. 10,731,181
2019	U.S. Patent No. 10,745,716
2023	Krebber, H. and Silver, P.A., "Directing Proteins to Nucleus by Fusion to Nuclear Localization Signal Tags," <i>Methods in Enzymology</i> 327: 283-296 (2000)
2026	Prov. Appl. No. 61/734,256, filed December 6, 2012
2031	Jinek, M., <i>et al.</i> , "A Programmable Dual-RNA-Guided DNA Endonuclease in Adaptive Bacterial Immunity," <i>Science</i> 337(6096):816-821, with Supplementary Information (2012)
2221	Lange, A., <i>et al.</i> , "Classical Nuclear Localization Signals: Definition, Function, and Interaction with Importin $\alpha$ ," <i>J. Biol. Chem.</i> 282(8): 5101–5105 (2007)
2348	Doudna, J.A., and Lorsch, J.R., "Ribozyme catalysis: not different, just worse," <i>Nature Structure &amp; Molecular Biology</i> 12(5):395-402 (2005)
2400	Decision on Motions 37 C.F.R. § 41.125(a), <i>The Regents of the University of California v. The Broad Institute, Inc.</i> , Patent Interference No. 106,115, Paper 877, (September 10, 2020)
2543	Declaration of Yannick Doyon, Ph.D.
2548	Declaration of Samuel Sternberg, Ph.D.
2549	Declaration of Scott Bailey, Ph.D.
2550	van der Aa, M.A.E.M., <i>et al.</i> , "The Nuclear Pore Complex: The Gateway to Successful Nonviral Gene Delivery," <i>Pharmaceutical Research</i> 23(3): 447- 459 (2006)
2560	Declaration 37 C.F.R. § 41.203(b), Count and claims of the parties, <i>The Regents of the University of California v. The Broad Institute, Inc.</i> , Patent Interference No. 106,115, Paper 1, (June 24, 2019)
2561	Declaration 37 C.F.R. § 41.203(b), Count and claims of the parties, <i>The Regents of the University of California v. ToolGen, Inc.</i> , Patent Interference No. 106,127, Paper 1, (December 14, 2020)
2562	Declaration 37 C.F.R. § 41.203(b), Count and claims of the parties, <i>The Broad Institute, Inc. v. ToolGen, Inc.</i> , Patent Interference No. 106,126, Paper 1, (December 14, 2020)
2585	U.S. Appl. No. 15/456,204 with current claims appended
2655	Deposition Transcript of Paula Cannon, Ph.D., (March 28, 2022)

1                    **APPENDIX B: STATEMENT OF MATERIAL FACTS**

2                    **CVC’S STATEMENT OF MATERIAL FACTS AND SIGMA’S RESPONSES**

3    **1.**        CVC’s half of Count 1 aligns with the scope of the counts in multiple ongoing  
4 interferences, including interferences between: CVC and Broad (Interference No. 106,115), Ex.  
5 2560 at 12-13; CVC and ToolGen (Interference No. 106,127), Ex. 2561 at 5-6; and Broad and  
6 ToolGen (Interference No. 106,126), Ex. 2562 at 12-13.

7                    **Response: Denied**

8    **2.**        In each of these Interferences 106,115, 106,127 and 106,126, both halves of the count  
9 recite single-guide (“sgRNA”) CRISPR-Cas9 systems, whether or not the parties have claims  
10 that are generic as to the format of the guide RNA (i.e., “generic-guide” claims). Ex. 2560, 12-  
11 13; Ex. 2561 at 5-6; Ex. 2562, 12-13; *see* Ex. 2400, 31-33.

12                    **Response: Admitted**

13    **3.**        CVC has involved claims directed to both generic guide RNA and single-guide RNA  
14 embodiments. *See, e.g.*, CVC Application Nos. 15/947,700; 15/947,718; 15/981,808; 15/981,809  
15 (reciting CRISPR-Cas9 systems in eukaryotic cells that are not limited to single-molecule RNA).

16                    **Response: Denied**

17    **4.**        Count 1 in this Interference is defined as CVC Application 15/981,807, claim 156 or  
18 Sigma Application 15/456,204 claim 31 (colloquially referred to as a “McKelvey count”). *See*  
19 Paper 1, Declaration of Interference, 5.

20                    **Response: Admitted**

21    **5.**        CVC claim 156 recites “a single molecule DNA-targeting RNA, or a nucleic acid  
22 comprising a nucleotide sequence encoding said single molecule DNA-targeting RNA” and then  
23 goes on to define the three essential components of a single-molecule guide RNA. *See* Paper 1,  
24 Declaration of Interference, 5.

1           **Response: Denied**

2    6.       Sigma’s claim 31 recites “a guide RNA or DNA encoding the guide RNA, wherein the  
3    guide RNA comprises a first region that is complementary to a target site in the chromosomal  
4    sequence . . . and a second region that interacts with the CRISPR-Cas type-II protein, and  
5    wherein the guide RNA comprises a crRNA and a tracrRNA.” See Paper 1, Declaration of  
6    Interference, 6.

7           **Response: Denied**

8    7.       The PTAB construed the term “guide RNA” in the ’115 Interference to refer to sgRNA.  
9    Ex. 2400, 31-33.

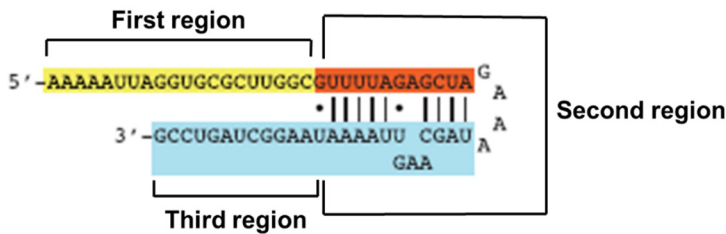
10          **Response: Denied**

11   8.       Claim 33 of Sigma’s ’204 Application adds the limitation “wherein the guide RNA is a  
12   single molecule.” Ex. 2585, 104.

13          **Response: Admitted**

14   9.       Sigma’s P1 application defines “guide RNA” as comprising a first region, “a second  
15   internal region that forms a stem loop structure,” and a third region. Ex. 2026, ¶¶ [0021], [0019].

16   These regions can be illustrated according to the figure below:



17  
18   Jinek, *Science* 2012 Fig. 5A (annotated) (cited in Sigma’s ’204 application). Ex. 2549, ¶ 99. The  
19   “loop” of claim 31 refers to the linker loop in the stem-loop region shown in purple in the  
20   diagram above, which joins the crRNA and the tracrRNA. Ex. 2549, ¶¶ 98-99, 101.

21          **Response: Denied**

1 **10.** Sigma’s ’204 application discloses data from a single experiment: a gel that has lanes for  
2 both single-guide RNA and dual-guide RNA molecules. Ex. 2585, 11-75; Ex. 2549, ¶103.

3 **Response: Denied**

4 **11.** CVC’s co-pending motion for priority benefit (CVC’s Substantive Motion 1) supports  
5 priority benefit based on CVC’s half of the count (claim 156 of CVC’s ’807 Application).

6 **Response: Denied**

7 **12.** Application No. 61/652,086 (“P1”), filed on May 25, 2012, lists Martin Jinek, Jennifer  
8 Doudna, Emmanuelle Charpentier, and Krzysztof Chylinski as co-inventors. Ex. 2009, 195.

9 **Response: Admitted**

10 **13.** Application No. 61/716,256 (“P2”), filed on October 19, 2012, lists Jinek, Doudna,  
11 Charpentier, Chylinski, and James Harrison Doudna Cate as co-inventors. Ex. 2010, 277.

12 **Response: Admitted**

13 **14.** CVC’s P1 describes CRISPR-Cas systems comprising a) a Cas9 protein and b) a single  
14 molecule DNA-targeting RNA. Ex. 2009, ¶¶ [00248]-[00251], Figs. 1-3; Ex. 2543, ¶¶ 90-242.

15 **Response: Admitted**

16 **15.** CVC’s P1 describes a sgRNA comprising i) a targeter RNA capable of hybridizing with a  
17 target sequence in the target DNA and ii) an activator-RNA capable of hybridizing with the  
18 targeter RNA to form a double-stranded duplex, wherein the activator-RNA and the targeter-  
19 RNA are covalently linked to one another with intervening nucleotides. Ex. 2009, ¶¶ [0079],  
20 [00119], [00248], Figs. 1, 3, 9; Ex. 2543, ¶¶ 90-95, 106-108, 175-179, 223.

21 **Response: Admitted**

22 **16.** CVC’s P1 describes a sgRNA capable of forming a complex with Cas9 and thereby  
23 targeting the Cas9 protein to the target DNA molecule. Ex. 2009, ¶¶ [0046], [0048], [0076],  
24 [0089], [00155]-[00156], [00248]-[00251], Figs. 1, 3; Ex. 2543, ¶¶ 90-95, 110-112, 180, 223.

1           **Response: Admitted**

2   **17.**    CVC’s P1 describes CRISPR-Cas9 systems capable of cleaving or editing target DNA or  
3   modulating transcription of at least one gene encoded by the target DNA. Ex. 2009, ¶¶ [00155]-  
4   [00159], [00248]-[00251], Figs. 3, 4; Ex. 2543, ¶¶ 90-95, 113-114, 180, 223.

5           **Response: Denied**

6   **18.**    CVC’s P1 describes target cells including a fish, a human, and a fruit fly cell, and that a  
7   target cell may be “embryonic.” Ex. 2009, ¶¶ [00165], [00216], [00218], [00050]-[00052],  
8   [00174].

9           **Response: Denied**

10   **19.**    CVC’s P1 describes making and using a single-molecule DNA-targeting RNA and a  
11   Cas9 RNA. Ex. 2009, ¶¶ [00173], [00248]; Ex. 2543, ¶¶ 90-95, 100, 170-173, 222.

12           **Response: Denied**

13   **20.**    CVC’s P1 describes that Cas9 can be delivered into a eukaryotic cell “as a polypeptide,”  
14   as a nucleic acid encoding Cas9, or in a pre-formed RNP complex. Ex. 2009, ¶¶ [00120],  
15   [00126]-[00128], [00167]-[00172], [00177]-[00178]; Ex. 2543, ¶¶ 92, 96-99, 115, 132-135, 140.

16           **Response: Denied**

17   **21.**    CVC’s P1 describes that the sgRNA can be delivered into a eukaryotic cell “directly as  
18   RNA” or as a nucleic acid “comprising a nucleotide sequence encoding a subject DNA-targeting  
19   RNA.” Ex. 2009, ¶¶ [00120], [00167], [00170]-[00173], [00177]; Ex. 2543, ¶¶ 92, 96-99, 137-  
20   140.

21           **Response: Denied**

22   **22.**    CVC’s P1’s working example describes incubating a recombinant Cas9 protein with the  
23   sgRNA to make an RNP complex. Ex. 2009, ¶¶ [00248]-[00251]; Ex. 2543, ¶¶ 92, 96-99, 137-  
24   140.



1           **Response: Admitted**

2   **23.**    CVC’s P1’s working example describes a sgRNA complexed with a Cas9 protein  
3   cleaving a target DNA. Ex. 2009, ¶¶ [00248]-[00251], Fig. 3A; Ex. 2543, ¶¶ 92, 96-99, 137-140.

4           **Response: Admitted**

5   **24.**    CVC’s P1 describes microinjection as a method of delivering Type II CRISPR-Cas9 into  
6   a cell. Ex. 2009, ¶¶ [0039], [00154], [00173]-[00175]; Ex. 2543, ¶¶ 141-146, 225.

7           **Response: Denied**

8   **25.**    By May 25, 2012, microinjecting protein, RNA, or RNPs into eukaryotic cells were well-  
9   known, routine laboratory techniques. Ex. 2009, ¶ [00173]; Ex. 2543, ¶¶ 66-72.

10          **Response: Denied**

11   **26.**    CVC’s P1 describes transfection as a method for delivering Type II CRISPR-Cas9  
12   systems into a cell. Ex. 2009, ¶¶ [00129], [0039], [00154], [00173-175], [00177]; Ex. 2543, ¶¶  
13   199-200.

14          **Response: Denied**

15   **27.**    By May 25, 2012, transfecting proteins, RNA, and RNPs into eukaryotic cells human cell  
16   lines were well-known, routine laboratory techniques. Ex. 2009, ¶ [00173]; Ex. 2543, ¶¶ 73-82.

17          **Response: Denied**

18   **28.**    By May 25, 2012, the art disclosed that a PAM must be adjacent to the target sequence  
19   for Type II CRISPR-Cas9 systems to cleave target DNA. Ex. 2543, ¶¶ 54-64, 249-259.

20          **Response: Denied**

21   **29.**    CVC’s P1 discloses a PAM sequence adjacent to the target in Target DNA A (“GGG”),  
22   Target DNA B (“GGG”), and Target DNA C (“TGG”). Ex. 2009, Fig. 3C; Ex. 2543, ¶¶ 249-259.

23          **Response: Denied**

1 **30.** CVC's P1 describes "replac[ing] a codon with a codon encoding the same amino acid."  
2 Ex. 2009, ¶ [0033]; Ex. 2543, ¶¶ 190, 285-289.

3 **Response: Denied**

4 **31.** CVC's P1 describes peptide that can be added to Cas9, including a polypeptide that  
5 facilitates traversing an organelle membrane. Ex. 2009, ¶ [00115]; Ex. 2543, ¶¶ 120-121, 277-  
6 284.

7 **Response: Denied**

8 **32.** All of the disclosures in CVC's P1 are in P2. Ex. 2009; Ex. 2010; Ex. 2543, ¶¶ 243-245.

9 **Response: Admitted**

10 **33.** CVC's P2 describes PAMs and cites Saprunauskas (Ex. 2132), Deveau (Ex. 2125),  
11 Mojica (Ex. 2127), Makarova (Ex. 2130), and Wiedenheft (Ex. 2134), which discuss PAMs in  
12 CRISPR-Cas systems. Ex. 2010, ¶¶ [00103], [00350]-[00352], [00359]; Ex. 2543, ¶¶ 243-245.

13 **Response: Denied**

14 **34.** CVC's '859 application was filed within 12 months of the filing dates of P1 and P2, and  
15 makes specific reference to CVC's P1 and P2 applications. Ex. 2015, 5.

16 **Response: Admitted**

17 **35.** CVC's '504 application was filed during the '859 application's pendency and makes  
18 specific reference to CVC's '859, P1, and P2 applications. Ex. 2006, 4-5.

19 **Response: Admitted**

20 **36.** CVC's '604 application was filed during the '504 application's pendency and makes  
21 specific reference to CVC's '504, '859, and P1 and P2 applications. Ex. 2007, 356-360.

22 **Response: Admitted**

23 **37.** Target DNA A, disclosed in Figure 3C of CVC's P1 and P2, is a non-natural target and  
24 P1 and P2 disclose Target DNA A as including a PAM. Ex. 2009, Fig. 3C; Ex. 2543, ¶¶ 255-259.

1           **Response: Denied**

2   **38.**    CVC's P1 describes and enables modification of a chromosomal sequence in a eukaryotic  
3 cell by integrating a donor sequence, which occurs when the cell's own homology-directed repair  
4 process repairs DNA cut by a nuclease, whether a sgRNA-CRISPR-Cas9, a ZFN, or a TALEN.  
5 Ex. 2009, ¶¶ [0058], [0059], [00157], [00189]-[00193], Fig. 4, claims 77, 88, 99; Ex. 2543, ¶¶  
6 126-128, 303.

7           **Response: Denied**

8   **39.**    The differences between claim 1 of the '181 or '716 patent and Sigma's half of Count 1,  
9 are that the claims specify which Cas9 protein to use (from *S. pyogenes*), where to locate the  
10 DNA-targeting region within the guide RNA (at the 5' end), and which NLS to use (C-terminal  
11 SEQ ID NO: 1 or SEQ ID NO: 2). Ex. 2017, 71:34-72:39; Ex. 2019, 71:14-51; Ex. 2549, ¶¶9,  
12 30.

13           **Response: Denied**

14   **40.**    Jinek 2012 discloses *in vitro* experiments that used *S. pyogenes* Cas9 to cleave target  
15 DNA, including GFP, a sequence from a eukaryote. Ex. 2031, Figs. 1-5; Ex. 2549, ¶¶47, 57.

16           **Response: Denied**

17   **41.**    Jinek 2012 discloses *in vitro* experiments using guide RNAs comprising a DNA-targeting  
18 region at the 5' end that base pairs with a target site in the chromosomal sequence. Ex. 2031,  
19 Figs. 1E, 3C, 5B; Ex. 2549, ¶¶34-37.

20           **Response: Denied**

21   **42.**    The natural location for the DNA-targeting region of a guide RNA in a CRISPR system  
22 is at the 5' end. Ex. 2031, 818; Ex. 2549, ¶34.

23           **Response: Denied**

1 **43.** Krebber 2000 discloses methods of using the NLS listed as SEQ ID NO: 1. Ex. 2023,  
2 285; Ex. 2549, ¶¶39.

3 **Response: Denied**

4 **44.** Krebber 2000 discloses attaching an NLS at either the N-terminus or C-terminus of the  
5 tagged protein. Ex. 2023, 289-290; Ex. 2549, ¶39.

6 **Response: Denied**

7 **45.** Lange 2007 discloses methods of using the NLS listed as SEQ ID NO: 2. Ex. 2221, 3;  
8 Ex. 2549, ¶39.

9 **\*Sigma did not respond to CVC's MF 45, and therefore this MF should be deemed**  
10 **admitted.\***

11 **46.** Before December 2012, the SV40 NLS was the most commonly used NLS peptide for  
12 tagging proteins. Ex. 2550, 451-452; Ex. 2023, 285; Ex. 2549, ¶39.

13 **Response: Denied**

14 **47.** Before December 2012, the SV40 NLS was “the model” for NLSs. Ex. 2348, 478; Ex.  
15 2549, ¶39.

16 **Response: Denied**

17 **48.** All the disclosures of CVC's P1 application appear in CVC's P2 application, and in  
18 CVC's U.S. Provisional Application No. 61/757,640, as well as in CVC's U.S. Appl. No.  
19 13/842,859. Ex. 2009; Ex. 2010; Ex. 2011; Ex. 2015; Ex. 2543, ¶¶ 219, 243-245, 316.

20 **Response: Denied**

21 **49.** Expressing a dual molecule guide RNA would require multiple promoters. Ex. 2549,  
22 ¶100.

23 **Response: Denied**

1 **50.** Sigma has not requested de-designating any claims from this proceeding. *See* Order  
2 Authorizing Motions and Setting Times 37 C.F.R. §§ 104(c) and 121.

3 **Response: Denied**

4

1           **CVC’S RESPONSES TO SIGMA’S STATEMENT OF MATERIAL FACTS**

2   **51.**    CVC’s Proposed Count 2 includes CVC Claim 156, which (for purposes of this  
3 interference) is substantively the same as Claim 156 of Sigma’s Proposed Count 2.

4           **Response: Denied.**

5   **52.**    In CVC’s Motion 3, CVC does not contend that changing the count would better conform  
6 to CVC’s proofs of invention.

7           **Response: Admitted.**

8   **53.**    Sigma’s proofs of invention, as set forth in Sigma P1, are directed to the use of both  
9 sgRNA and dgRNA in a CRISPR-Cas9 system. Ex. 1080 ¶¶ 76-79.

10           **Response: Admitted that Sigma’s P1, discloses both sgRNA and dgRNA in a**  
11 **CRISPR-Cas9 system, otherwise denied.**

12   **54.**    Sigma’s proofs of invention, as set forth in Sigma P1, are not identical for both types of  
13 guide RNAs (sgRNA and dgRNA). *Id.*

14           **Response: CVC does not have adequate information regarding Sigma’s proofs of**  
15 **invention to admit or deny.**

16   **55.**    In opposition proceedings before the European Patent Office (“EPO”), CVC’s exclusive  
17 licensee Intellia Therapeutics (via Intellia’s European Patent Attorney George Schlich)  
18 challenged Sigma’s European applications related to CRISPR-Cas9. Exs. 1085-86.

19           **Response: Admitted that attorney George Schlich participated in EPO proceedings**  
20 **involving Sigma’s European patent applications, otherwise denied.**

21   **56.**    In EPO proceedings, Intellia criticized Sigma’s experimental data using sgRNA (Lanes  
22 B, C, & D), arguing that Sigma’s lack of PCR confirmations in those lanes demonstrated lack of  
23 successful donor integration via HDR. *Id.*

1           **Response: Admitted that attorney George Schlich participated in EPO proceedings**  
2 **involving Sigma’s European patent applications, otherwise denied.**

3 **57.** In its pertinent disclosure, CVC P3 demonstrated cleavage in eukaryotic cells using only  
4 a sgRNA. Ex. 1080 ¶ 77.

5           **Response: Admitted that CVC P3 demonstrated cleavage in eukaryotic cells using a**  
6 **sgRNA.**

7 **58.** In its pertinent disclosure, Broad P1 demonstrated cleavage in eukaryotic cells using only  
8 a sgRNA. *Id.*

9           **Response: Denied.**

10 **59.** In its pertinent disclosure, ToolGen P1 demonstrated cleavage in eukaryotic cells using  
11 only a sgRNA. *Id.*

12           **Response: Admitted that ToolGen’s P1 discloses only sgRNA.**

13 **60.** The term “loop” is recited nowhere in Sigma’s patent claims, including Sigma claim 31.  
14 *Id.* ¶ 81. A2-12.

15           **Response: Admitted that the word “loop” does not appear in Sigma’s patent claims,**  
16 **but denied that Sigma’s claim 31, as properly construed, covers a guide RNA**  
17 **lacking a loop of a “stem loop structure.”**

18 **61.** CVC’s inserted “Fig. 5A [sic] (annotated)” figure is adapted from Jinek (2012)’s  
19 illustration of a “chimera A” sgRNA discussed by Jinek (Ex. 2031, Fig. 5B), and is not a figure  
20 from Sigma P1. *Id.*

21           **Response: Admitted.**

22 **62.** Sigma P1 ¶ [0023] explains that the guide RNA—including a “stem loop”—can be a  
23 *dual-molecule* guide RNA. *Id.*

24           **Denied.**

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**CVC’S ADDITIONAL STATEMENT OF MATERIAL FACTS IN REPLY**

**63.** Sigma’s only purported difference between its single-guide and dual-molecule proofs is that it has PCR evidence confirming the dual-molecule experiments that it does not have for the single-guide experiments. Paper 708, Sigma Opp. at 7:11-24.

**64.** Sigma also does not allege that its alleged date of invention would be earlier for its dual-guide proofs than its single-guide proofs. Paper 708, Sigma Opp at 7:11-24.

**65.** Ex. 2585, ¶¶ [0071]-[0072] describes a single molecule of RNA in which the crRNA and tracrRNA are physically linked by a loop to form a “stem loop structure.”

**66.** There is no physically possible guide RNA structure that is both (a) a dual-molecule guide RNA and (b) a guide RNA with a stem-loop in the second region as defined by Sigma’s specification. *See* Ex. 1082, 85:4-86:6 (explaining that the only way a guide RNA can comprise two molecules is if it does *not* have an intact stem loop in the second region).

**67.** Sigma’s expert offered no analysis of the structure of the single-guide or dual-guide RNA purportedly disclosed in Sigma’s involved application. Ex. 2655, 119:16-127:13.



**CERTIFICATE OF SERVICE**

I hereby certify that the foregoing **JUNIOR PARTY’S REPLY MOTION 3**, is being filed via the Interference Web Portal by 8:00 PM Eastern Time on April 7, 2022, pursuant to the Order Authorizing Motions and Setting Times (“Order”; Paper 30), and thereby served on the attorney of record for the Senior Party pursuant to ¶ 105.3 of the Standing Order. Pursuant to the Order, the foregoing was also served via email by 11:00 PM Eastern Time on counsel for the Senior Party at:

Brenton R. Babcock  
Dan Liu, Ph.D.  
LOEB & LOEB LLP  
10100 Santa Monica Blvd., Ste. 2200  
Los Angeles, CA 90067  
bbabcock@loeb.com  
dliu@loeb.com  
BoxSigma132@loeb.com

Benjamin J. Sodey  
SIGMA-ALDRICH CORP.  
3050 Spruce St.  
Saint Louis, MO 63103  
benjamin.sodey@milliporesigma.com

Benjamin I. Dach, Ph.D.  
LOEB & LOEB LLP  
345 Park Ave.  
New York, NY 10154  
bdach@loeb.com

STERNE, KESSLER, GOLDSTEIN & FOX P.L.L.C

/Eldora L. Ellison/  
Eldora L. Ellison, Ph.D., Esq.  
Lead Attorney for UC and UV  
Registration No. 39,967

Date: April 7, 2022

STERNE, KESSLER, GOLDSTEIN & FOX P.L.L.C.  
1100 New York Avenue, NW  
Washington, DC 20005-3934