

UNITED STATES PATENT AND TRADEMARK OFFICE

BEFORE THE PATENT TRIAL AND APPEAL BOARD

MERCK SHARP & DOHME CORP.
Petitioner

v.

GENENTECH, INC. AND CITY OF HOPE
Patent Owners

U.S. Patent No. 6,331,415

“Methods of Producing Immunoglobulins, Vectors and
Transformed Host Cells for Use Therein”

Inter Partes Review No. 2017-00047

**PETITION FOR *INTER PARTES* REVIEW OF U.S. PATENT NO. 6,331,415
UNDER 35 U.S.C. §§ 311-319 AND 37 C.F.R. §§ 42.100 *et seq.***

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EXHIBIT LIST

Exhibit No.	Exhibit Description	Abbreviation
1001	U.S. Patent No. 6,331,415	The '415 patent
1002	U.S. Patent No. 4,495,280	Bujard, or the Bujard Patent
1003	Riggs and Itakura, <i>Synthetic DNA and Medicine</i> , American Journal of Human Genetics, 31:531-538 (1979)	Riggs & Itakura
1004	Southern and Berg, <i>Transformation of Mammalian Cells to Antibiotic Resistance with a Bacterial Gene Under Control of the SV40 Early Region Promoter</i> , Journal of Molecular and Applied Genetics, 1:327-341 (1982)	Southern
1005	U.S. Patent No. 4,237,224	Cohen & Boyer, or the Cohen & Boyer patent
1006	Declaration of Margaret H. Baron, M.D., Ph.D., in Support of Merck Sharp & Dohme Corp.'s Petition for <i>Inter Partes</i> Review of U.S. Patent No. 6,331,415	Baron Decl.
1007	U.S. Patent No. 4,816,657	The Cabilly I patent
1008	'415 patent reexamination, Office Action dated 2/16/07	Office Action (2/16/07)
1009	'415 patent reexamination, Owners' Resp. dated 11/25/05	Owners' Resp. (11/25/05)
1010	'415 patent reexamination, Owners' Resp. (5/21/07)	Owners' Resp. (5/21/07)

Exhibit No.	Exhibit Description	Abbreviation
1011	'415 patent reexamination, Office Action dated 9/13/05	Office Action (9/13/05)
1012	U.S. Patent No. 4,816,397	The Boss patent
1013	'415 patent file history, Paper 17	
1014	'415 patent file history, Paper 14	
1015	'415 patent file history, Paper 18	
1016	'415 patent reexamination, Office Action dated 8/16/06	Office Action (8/16/06)
1017	'415 patent reexamination, Office Action dated 2/25/08	Office Action (2/25/08)
1018	U.S. Patent No. 4,399,216	Axel, or the Axel patent
1019	U.S. Patent No. 5,840,545	Moore, or the Moore patent
1020	Rice and Baltimore, <i>Regulated Expression of an Immunoglobulin K Gene Introduced into a Mouse Lymphoid Cell Line</i> , Proceedings of the National Academy of Sciences USA, 79:7862-7865 (1982)	Rice & Baltimore
1021	Ochi et al., <i>Transfer of a Cloned Immunoglobulin Light-Chain Gene to Mutant Hybridoma Cells Restores Specific Antibody Production</i> , Nature, 302:340-342 (1983)	Ochi (I)
1022	'415 patent reexamination, Owners' Resp. dated 10/30/06	Owners' Resp. (10/30/06)
1023	'415 patent reexamination, Owners' Resp.	Owners' Resp.

Exhibit No.	Exhibit Description	Abbreviation
	dated 6/6/08	(6/6/08)
1024	'415 patent reexamination, Appeal Brief	Appeal Brief
1025	'415 patent reexamination, Notice of Intent to Issue Ex Parte Reexamination Certificate	NIRC
1026	'415 reexamination, Ex Parte Reexamination Certificate	Reexam Cert.
1027	T.J.R. Harris, <i>Expression of Eukaryotic Genes in E. Coli</i> , in Genetic Engineering 4, 127-185 (1983)	Harris
1028	'415 patent reexamination, Declaration of Dr. Timothy John Roy Harris under 37 C.F.R. § 1.132	Harris Decl.
1029	Kabat et al., Sequences of Proteins of Immunological Interest (1983) (excerpt)	Kabat
1030	Cohen, <i>Recombinant DNA: Fact and Fiction</i> , Science, 195:654-657 (1977)	Cohen
1031	Oi et al., <i>Immunoglobulin Gene Expression in Transformed Lymphoid Cells</i> , Proceedings of the National Academy of Sciences USA, 80:825-829 (1983)	Oi
1032	European Patent Application Publication No. 0044722 A1, published 1/27/82	Kaplan
1033	U.S. Patent No. 4,487,835	
1034	U.S. Patent No. 4,371,614	
1035	U.S. Patent No. 4,762,785	
1036	U.S. Patent No. 4,476,227	

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1038	U.S. Patent No. 4,396,601	
1039	Milstein, <i>Monoclonal Antibodies from Hybrid Myelomas</i> , Proceedings of the Royal Society of London, 211:393-412 (1981)	
1040	Ochi et al., <i>Functional Immunoglobulin M Production after Transfection of Cloned Immunoglobulin Heavy and Light Chain Genes into Lymphoid Cells</i> , Proceedings of the National Academy of Sciences USA, 80:6351-6355 (1983)	
1041	<i>MedImmune, Inc. v. Genentech, Inc.</i> , No. 03-02567 (C.D. Cal. Aug. 17, 2007), Expert Report of E. Fintan Walton	
1042	'415 patent reexamination, Request for Reconsideration and/or Petition Under 37 C.F.R. § 1.183 dated 5/15/09	
1043	Feldman et al., <i>Lessons from the Commercialization of the Cohen-Boyer Patents: The Stanford University Licensing Program</i> , in Intellectual Property Management in Health and Agricultural Innovation: A Handbook of Best Practices, 1797-1807 (2007)	
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Exhibit No.	Exhibit Description	Abbreviation
1046	'415 patent reexamination, Declaration of Dr. E. Fintan Walton under 37 C.F.R. § 1.132	
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1048	Stipulation and order of dismissal in <i>MedImmune v. Genentech</i> , No. 03-02567 (C.D. Cal.)	
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1056	Stipulation of dismissal in <i>Eli Lilly and ImClone Systems LLC v. Genentech</i> , No. 13-CV-7248 (C.D. Cal.)	
1057	Complaint in <i>Bristol-Myers Squibb v.</i>	

Exhibit No.	Exhibit Description	Abbreviation
	<i>Genentech</i> , No. 13-CV-5400 (C.D. Cal.)	
1058	Stipulation of dismissal in <i>Bristol-Myers Squibb v. Genentech</i> , No. 13-CV-5400 (C.D. Cal.)	

I. INTRODUCTION

Merck Sharp & Dohme Corp. (“Petitioner” or “Merck”) requests *inter partes* review (“IPR”) pursuant to 35 U.S.C. §§ 311-319 and 37 C.F.R. §§ 42.100 *et seq.* of claims 1-4, 11, 12, 14, 18-20, and 33 (“the challenged claims”) of U.S. Patent No. 6,331,415 (“the ’415 patent,” Ex. 1001), which issued on December 18, 2001 to Cabilly *et al.* and is assigned to Genentech, Inc. and City of Hope (“Owners”). A petition for *inter partes* review must demonstrate “a reasonable likelihood that the petitioner would prevail with respect to at least one of the claims challenged in the petition.” 35 U.S.C. § 314(a). This Petition meets this threshold for the reasons outlined below.

In this Petition, Merck asserts the same grounds of unpatentability upon which the Board has already instituted review of the challenged claims of the ’415 patent in *Mylan Pharmaceuticals, Inc. v. Genentech, Inc., et al.*, IPR2016-00710 (the “Mylan IPR”); *see* Paper 13 (Institution of Inter Partes Review, 37 C.F.R. §42.108) (Sept. 8, 2016), at 2. For the exact same reasons previously considered by the Board, on the exact same trial schedule, Merck respectfully seeks to join the Mylan IPR. This Petition does not add to or alter any arguments that have already been considered by the Board, and this Petition does not seek to expand the grounds of unpatentability that the Board has already instituted in the Mylan IPR. Accordingly, and as explained below, there exists a reasonable likelihood that

Merck will prevail in demonstrating unpatentability of at least one of the challenged claims based on teachings set forth in the references presented in this Petition.

Because this Petition is filed within one month of the institution of the Mylan IPR, and because this Petition is accompanied by a Motion for Joinder, this Petition is timely and proper under 35 U.S.C. § 315(c).

II. REQUIREMENTS FOR INTER PARTES REVIEW

A. Grounds for Standing (37 C.F.R. § 42.104(a))

Pursuant to 37 C.F.R. § 42.104(a), Petitioner certifies that the '415 patent is available for IPR and that Petitioner is not barred or estopped from requesting an IPR of the '415 patent on the grounds identified in this Petition. Petitioner further certifies that the prohibitions of 35 U.S.C. §§ 315(a)-(b) are inapplicable.

B. Identification of Challenge (37 C.F.R. § 42.104(b))

Petitioner requests that the Board cancel challenged claims 1-4, 11, 12, 14, 18-20, and 33 of the '415 patent. These are all of the claims that were challenged and all of the claims for which an IPR was instituted in the Mylan IPR. *See* Paper 13 (Institution of Inter Partes Review, 37 C.F.R. §42.108) at 2. Petitioner herein challenges the patentability of the same claims on the grounds as follows:¹

¹ Previously, claims 1-4, 9, 11, 12, 14-20, and 33 of the '415 patent had been challenged, on four asserted grounds of unpatentability (one anticipation ground

Ground 1. Claims 1, 3, 4, 11, 12, 14, 19, and 33 are obvious under § 103(a) over Bujard (Ex. 1002) in view of Riggs & Itakura (Ex. 1003); and

Ground 2. Claims 1, 2, 18, 20, and 33 are obvious under § 103(a) over Bujard in view of Southern (Ex. 1004).

The challenged claims of the '415 patent purport to cover recombinant DNA processes and associated compositions for making immunoglobulins (or

and three separate obviousness grounds), and the Board instituted an IPR of claims 1-4, 11, 12, 14, 18-20, and 33 of the '415 patent based on two of the asserted obviousness grounds. *See Sanofi-Aventis U.S. LLC, et al. v. Genentech, Inc., et al.*, IPR2015-01624 (the “Sanofi IPR”), Paper 15 (Institution of Inter Partes Review, 37 C.F.R. §42.108) (Feb. 5, 2016), at 22-23. A separate IPR petition filed by Petitioner Genzyme Corporation (IPR2016-00460) was instituted on the same grounds and joined to the Sanofi IPR. *See IPR2016-00460*, Paper 12; Sanofi IPR, Paper 35 (March 28, 2016). In the Mylan IPR, Mylan challenged the then-instituted claims only on the two previously instituted obviousness grounds and the Mylan IPR was instituted on those two grounds. *See Mylan IPR*, Paper 13 (Institution of Inter Partes Review, 37 C.F.R. §42.108) (Sept. 8, 2016) at 13-14. Petitioner herein challenges the patentability of these same challenged claims on the same two, thrice previously instituted, grounds.

antibodies) in “host” cells that are genetically engineered to contain the two DNA sequences encoding the heavy and light chain polypeptides necessary for the cell to make an immunoglobulin. The generally applicable techniques employed by the ’415 patent inventors were already disclosed and commonly used in the prior art, including the Bujard patent. This reference was not substantively considered by the PTO during prosecution or reexamination of the ’415 patent. Moreover, Bujard discloses the precise teachings that Owners previously argued were missing from the prior art: the introduction of “a plurality of” or “one or more” DNA sequences into a host cell—language which necessarily accommodates two DNA sequences, including the heavy and light chain sequences. Because Bujard also expressly identifies immunoglobulins as being among the types of proteins that can be made in host cells by their respective methods, Bujard, in view of the Riggs & Itakura reference, and separately, in view of Southern prior art reference, each render obvious the challenged claims of the ’415 patent.

Pursuant to 37 C.F.R. § 42.204(b), a detailed explanation of the precise relief requested for each challenged claim including where each element is found in the prior art and the relevance of the prior art reference is provided in Section V below. Additional explanation and support for each ground of unpatentability is set forth in the accompanying Declaration of Margaret H. Baron, M.D., Ph.D (Ex. 1006), in which Dr. Baron offers the same opinions as those set forth by Dr.

Jefferson Foote in connection with the Sanofi IPR and by Dr. Kathryn Calame in the Mylan IPR.

As noted *supra*, the Mylan IPR is premised on the same opinions. In its petition, Mylan sought to join the then-pending Sanofi IPR. In support of its own petition, Mylan re-submitted Dr. Foote's Declaration from the Sanofi IPR, *see* IPR2016-00710, Ex. 1006 (Declaration of Jefferson Foote, Ph.D., in Support of Sanofi And Regeneron's Petition for *Inter Partes* Review of U.S. Patent No. 6,331,415), and also retained its own expert, Dr. Calame, who "fully agree[s] with, and hereby adopt[s], the opinions set forth in the Foote Declaration." *See* IPR2016-00710, Ex. 1059 (Declaration of Kathryn Calame, Ph.D., in Support of Mylan Pharmaceuticals Inc.'s Petition for *Inter Partes* Review of U.S. Patent No. 6,331,415) at ¶16. The Sanofi IPR has since been terminated (along with the joined IPR2016-00460). *See* Paper 43 (Termination of the Proceeding, 37 C.F.R. § 42.73) (Sept. 2, 2016), at 1-2.

The Baron Declaration supporting this Petition does not alter or otherwise seek to supplement the opinions offered by Dr. Foote in the Sanofi IPR, on which Mylan currently relies, or the opinions offered by Dr. Calame, who adopted Dr. Foote's opinions. And Dr. Baron does not intend to offer opinions beyond those expressed by Dr. Foote in connection with the Sanofi IPR and adopted by Dr. Calame in the connection with the Mylan IPR. For at least the reasons set forth in

Merck's Motion for Joinder, filed concurrently herewith, Merck respectfully requests institution of trial on the unpatentability grounds detailed below and joinder with the Mylan IPR.

III. RELEVANT INFORMATION REGARDING THE '415 PATENT

A. Brief Description of the Challenged Patent

The '415 patent issued on December 18, 2001, from Application No. 07/205,419 ("the '419 application"), filed on June 10, 1988. The '419 application has an earliest effective filing date under 35 U.S.C. § 120 of April 8, 1983, by virtue of a priority claim to Application No. 06/483,457, which issued as U.S. Patent No. 4,816,567 ("the Cabilly I patent," Ex. 1007). A reexamination certificate for the '415 patent issued on May 19, 2009, based on two separate reexamination requests filed on May 13 and December 23, 2005.

The '415 patent is directed to processes and related compositions for making immunoglobulins² (or fragments thereof) in host cells using recombinant DNA technology. Ex. 1001, 1:14-21, 3:53-67. Immunoglobulins are proteins (or "polypeptides") having a globular conformation that are produced by and secreted from cells of the immune system of vertebrates in response to the presence in the

² For purposes of this Petition, the claim term "immunoglobulin" is interchangeable with "antibodies," which the '415 patent defines as "specific immunoglobulin polypeptides." Ex. 1001, 1:23-24

body of a foreign substance, called an “antigen,” often a foreign protein or a foreign cell (such as a bacterium). *Id.* at 1:23-37; 16:38-39; Ex. 1006, Baron Decl.

¶ 31. Immunoglobulins bind to antigens to rid the body of the foreign invader. Ex. 1001, 1:26-31; Ex. 1006, Baron Decl. ¶ 31.

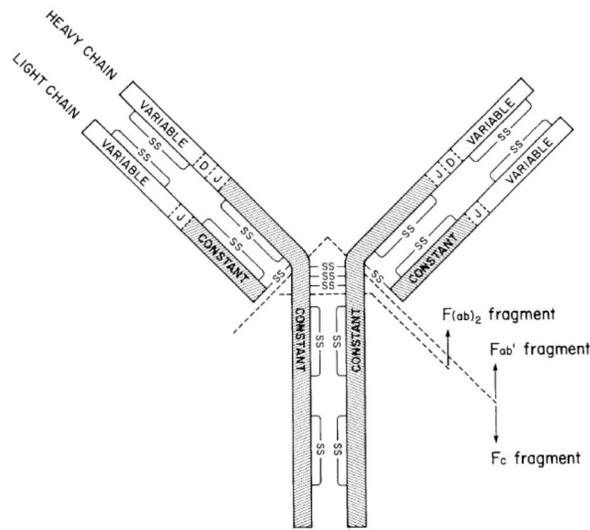


Fig. 1.

Ex. 1001, Fig. 1; Ex. 1006, Baron Decl. ¶ 31.

Most immunoglobulins are composed of two heavy chain polypeptides and two light chain polypeptides that are connected via disulfide bonds (represented above as –SS–) to form a four-chain “tetramer” with a highly specific and defined Y-shaped conformation that is required for antigen binding. Ex. 1001, Fig. 1 and 3:17-26; Ex. 1006, Baron Decl. ¶ 31. The heavy and light chains comprise segments referred to as the variable and constant regions. Ex. 1001, 3:42-59; Ex. 1006, Baron Decl. ¶ 32. The heavy chain and light chain are encoded by

separate DNA sequences or “genes.” Ex. 1001, 1:48-51; Ex. 1006, Baron Decl.

¶ 32. The nature of immunoglobulin structure and function as described above was well known in the prior art, as is evidenced by the discussion in the “Background of the Invention” in the ’415 patent. Ex. 1001 at 1:22-4:5; Ex. 1006, Baron Decl.

¶ 32.

The patent identifies a prior art method of making antibodies in hybridoma cells, which results in the production of a homogeneous antibody population that specifically bind to a single antigen, so called “monoclonal” antibodies. Ex. 1001 at 1:64-2:19. According to the patent, the use of recombinant DNA technology to make antibodies avoids the drawbacks of hybridoma production. *Id.* at 2:40-3:2

The recombinant DNA approach to making antibodies described in the patent, in short, proceeds as follows: (1) the genetic material encoding the heavy and light chains is identified and isolated (for example, from a hybridoma) (*id.* at 11:28-12:8; Ex. 1006, Baron Decl. ¶ 34); (2) the heavy and light chain DNA is introduced into suitable host cells by a process called “transformation,” which may be facilitated by first inserting the DNA into an expression vector³ that acts as a vehicle to introduce the foreign DNA into the host cell (Ex. 1001, 12:9-30;

³ Vectors that express inserted DNA sequences are called “expression vectors” in the patent, a term that is used interchangeably with “plasmid.” Ex. 1001, 8:16-22.

Ex. 1006, Baron Decl. ¶ 34); and (3) the host cells transcribe and translate the heavy and light chain DNA, a process called “expression,” to produce the heavy and light chain polypeptides (Ex. 1001, 12:31-33, 4:24-29; Ex. 1006, Baron Decl. ¶ 34). Host cells may either be microorganisms (for example, prokaryotic cells, such as bacteria) or cell lines from multicellular eukaryotic organisms, including mammalian cells. Ex. 1001, 8:41-56, 9:56-10:18.

The challenged claims of the '415 patent cover various aspects and components of the above-described recombinant production of immunoglobulins. All of the challenged claims (whether process or composition) require two genes: a first DNA sequence encoding the heavy chain and a second DNA sequence encoding the light chain. All of the challenged process claims require that the host cell express both DNA sequences to produce both heavy chain and light chain polypeptides (referred to as “co-expression” in the '415 patent and during reexamination⁴). Ex. 1009, Owners' Resp. (11/25/05), at 46. The heavy and light chain polypeptides are produced as “separate molecules” by virtue of their “independent expression.” Ex. 1001, claims 1, 33; Ex. 1022, Owners' Resp. (10/30/06), at 30 (“[T]he '415 patent requires that the transformed cell produce the immunoglobulin heavy and light chain polypeptides encoded by the two DNA

⁴ Ex. 1001, 12:50-51; Ex. 1008, Office Action (2/16/07), at 19.

sequences as separate molecules. This result stems from the requirement for independent expression of the introduced DNA sequences”).

Furthermore, the process claims also require assembly of the separate heavy and light chain polypeptides into an immunoglobulin tetramer. Ex. 1001, claim 1 (“A process for producing an immunoglobulin molecule”); Ex. 1009, Owners’ Resp. (11/25/05), at 46. This can occur inside of the host cell through its natural cellular machinery (“*in vivo*” assembly), which could then secrete the assembled immunoglobulin; or, if the host cell is unable to assemble the chains *in vivo*, the cell may be lysed and the separate chains assembled by chemical means (“*in vitro*” assembly). Ex. 1001, 12:50-55, claims 9 and 10; Ex. 1010, Owners’ Resp. (5/21/07), at 29, n.8.

B. Discussion of the File History and Related Proceedings in the PTO

The ’415 patent and the ’419 application have had an extended and extensive history in the PTO. The ’415 patent issued nearly thirteen-and-a-half years after its filing date and more than eighteen years after its priority filing date. During prosecution, the ’415 patent was involved in a decade-long interference proceeding (and related 35 U.S.C. § 146 action) with U.S. Patent No. 4,816,397, issued to Boss et al. (Ex. 1012). After the interference was resolved, prosecution of the ’415 patent continued until it issued. The ’415 patent was later the subject of an ex parte reexamination for four years, from May 13, 2005, to May 19, 2009.

1. Prosecution of the '419 application

The prosecution of the '419 application consisted largely of a series of restriction requirements by the PTO and claim cancellations and elections by Owners. *See generally* Ex. 1009, Owners' Resp. (11/25/05), at 8-10, 12-13. There were no prior art rejections of the pending claims. However, in an Information Disclosure Statement filed on September 18, 1991, Genentech characterized the Rice & Baltimore (Ex. 1020) prior art reference as “distinguishable from the instant claims in that the cells are not transformed with exogenous DNA encoding both of the heavy and light chains.” Ex. 1013, '415 patent file history, Paper 17, at 2 (emphasis in original).

1. Interference with the Boss Patent

On February 28, 1991, the Board of Patent Appeals and Interferences declared an interference between claims 1-18 of the Boss patent and then-pending claims 101-120 in the '419 application, which were copied from the Boss patent. Ex. 1014, '415 patent file history, Paper 14. The count was defined to be claim 1 of the Boss patent, which was identical to claim 101 of the '419 application (and which issued as claim 1 of the '415 patent). *Id.* at 4. The BPAI decided priority in favor of the senior party, Boss, holding that the inventors of the '415 patent had not established an actual reduction to practice before the Boss patent's British priority date. *Cabilly v. Boss*, 55 U.S.P.Q.2d 1238 (Bd. Pat. App. & Int. 1998). Priority of

invention was ultimately awarded to the inventors of the '415 patent on March 16, 2001, following settlement by the parties of an action instituted by Genentech under 35 U.S.C. § 146. Ex. 1015, '415 patent file history, Paper 18.

2. Ex Parte Reexamination of the '415 Patent

a. Rejections Over the Axel Patent

Over the course of reexamination, the PTO rejected the claims of the '415 patent in each of four office actions. *See* Exs. 1011, 1016, 1008, and 1017, '415 patent reexamination, Office Actions dated 9/13/2005, 8/16/2006, 2/16/2007, and 2/25/2008. Among the prior art relied upon by the PTO were U.S. Patent Nos. 4,399,216 (“Axel,” Ex. 1018) and 5,840,545 (“Moore,” Ex. 1019), Rice & Baltimore (Ex. 1020), and Ochi (I) (Ex. 1021). The PTO rejected the claims on a variety of grounds, including obviousness-type double patenting, anticipation, and obviousness. The OTDP rejections were in part based on (1) the claims of the Cabilly I patent, which were directed to chimeric⁵ heavy or light chains produced using recombinant DNA technology, in combination with (2) Axel, Rice & Baltimore or Ochi (I), alone or in combination with Moore. *E.g.*, Ex. 1008, Office

⁵ A “chimeric” chain has variable regions derived from one species of mammal, with constant portions derived from another species. *See* Ex. 1007, Cabilly I patent, 6:54-59 and claim 1.

Action (2/16/07), at 26-42. The obviousness rejections were based in part on the Moore patent either alone or in combination with the Axel patent. *Id.* at 12-14.

The PTO rejections relying on Axel were based on the Examiner's interpretation of Axel as disclosing the co-expression of heavy and light chains in a single host cell transformed with the respective DNA sequences. The invention of the Axel patent concerned "the introduction and expression of genetic informational material, i.e., DNA which includes genes coding for proteinaceous materials . . . into eucaryotic cells Such genetic intervention is commonly referred to as genetic engineering and in certain aspects involves the use of recombinant DNA technology." Ex. 1018, Axel, 1:12-21. Axel disclosed the transformation of eukaryotic (mammalian) host cells using a two-DNA system: "DNA I," which coded for a "desired proteinaceous material"⁶ that is "heterologous" to the host cell;⁷ and "DNA II," which coded for a protein that

⁶ A "desired proteinaceous material," or "protein of interest," is the protein that is sought to be isolated from the host cell after its production by the cell. Ex. 1010, Owner's Response (5/21/07), at 49; Ex. 1060, Baron Decl. ¶ 44.

⁷ A "heterologous" protein is a protein produced in a cell that does not normally make that protein or that is foreign to the cell, e.g., by genetically engineering the cell. Ex. 1060, Baron Decl. ¶ 44; Ex. 1001, 4:9-12, 4:33-41.

would act as a “selectable marker.”⁸ *Id.* at Fig. 1, 3:20-26, 8:56-62. Because DNA I and DNA II are present in a single vector “physically unlinked” to each other, (*id.* at 9:61-10:1; Fig. 1), the respective proteins encoded by DNA I and II would be independently expressed as separate molecules. Ex. 1006, Baron Decl. ¶ 44. The Axel patent identified “antibodies” as one of the preferred “proteinaceous materials” that could be made by the disclosed methods. Ex. 1018, Axel, 3:31-36, 2:61-66. In the first Office Action, the PTO characterized Axel as “demonstrat[ing] the predictability of expression of multiple heterologous proteins in a single host cell [and the] desirability of expressing immunoglobulins in mammalian host cells, and as intact (assembled) proteins.” Ex. 1011, Office Action (9/13/05), at 5.

The Examiner eventually entered a Final Office Action rejecting the claims in part over Axel, stating that the “Axel Abstract and definitions suggest cotransforming more than one desired gene for making proteinaceous materials

⁸ The function of a “selectable marker” is to permit scientists to identify which host cells have been transformed. Because it is not intended to be isolated or studied, it is not, strictly speaking, a protein “of interest” or a “desired” protein. Ex. 1009, Owners’ Response (11/25/05), at 34; Ex. 1060, Baron Decl. ¶ 44, n.3.

which include multimeric proteins.”⁹ Ex. 1017, Office Action (2/25/08), at 29; *see also id.* at 30 (“The Axel reference clearly encompasses one or more genes which encode one or more proteins.”). Moreover, the Examiner also found that Axel “teaches co-expression of two different proteins encoded by foreign DNA I and foreign DNA II in a single eukaryotic host cell.” *Id.* at 28. Because the proteins disclosed in the Axel patent included “multimeric proteins particularly ‘. . . interferon protein, antibodies, insulin, and the like,’” the Examiner concluded that “the Axel reference suggests expressing two immunoglobulin chains in a single eukaryotic host cell, since Axel discloses and claims encoding an antibody that necessarily possesses both light and heavy immunoglobulin chains.” Ex. 1008, Office Action (2/16/07), at 51.

Based on these rejections and others, all thirty-six of the ’415 patent claims stood finally rejected by the PTO over the prior art in the Office Action dated Feb. 25, 2008. Ex. 1017, Office Action (2/25/08), at 1.

⁹ A “multimeric” protein is a protein that is composed of more than one distinct polypeptide constituents or subunits. Ex. 1009, Owners’ Response (11/25/05), at 37. An immunoglobulin is a multimeric protein because it is composed of four distinct polypeptide subunits: two heavy chains and two light chains. Ex. 1022, Owners’ Response (10/30/06), at 33; Ex. 1060, Baron Decl. ¶¶ 72-74.

b. Owners' Arguments in Response to the Rejections

i. Owners Contrive a So-Called "Prevailing Mindset" before April 1983 that Only One Eukaryotic Protein of Interest Should be Produced in a Transformed Host Cell

In response to the rejections, Owners argued over the course of four responses and an appeal brief¹⁰ (relying on no fewer than seven Rule 132 declarations from technical experts) that none of the prior art disclosed the coexpression of both immunoglobulin heavy and light chains in a single host cell transformed with the genes encoding for both the heavy and light chains. Owners framed their specific arguments about the teachings of Axel with the general proposition that before April 1983, the so-called "prevailing mind-set" among persons of ordinary skill in the art (POSITA) was "that only one eukaryotic polypeptide of interest should be produced in a recombinant host cell." Ex. 1023, Owners' Resp. (6/6/08), at 6; *see also, e.g.*, Ex. 1024, Appeal Brief, at 33, 46 ("conventional 'one polypeptide at a time' approach" and "prevailing 'one polypeptide in a host cell' mindset"). This "prevailing mindset" would have led a POSITA "to break down a complex project, such as production of a multimeric eukaryotic protein, into more manageable steps (*e.g.*, produce each constituent

¹⁰ Exs. 1009, 1022, 1010, 1023, 1024, Owners' Responses, dated 11/25/05, 10/30/06, 5/21/07, and 6/6/08, and Appeal Brief, respectively.

polypeptide of the multimer in a separate host cell).” Ex. 1023, Owners’ Resp. (6/6/08), at 6-7. (As discussed below (at pages 21-25), this was decidedly not the prevailing mindset before April 1983: there were multiple prior art references teaching the expression of one or more genes in a single transformed host cell.)

This mindset, Owners argued, was reflected specifically in Axel. Ex. 1023, Owners’ Resp. (6/6/08), at 24-27. At most, according to Owners, Axel disclosed no more than producing either the heavy chain or light chain (or their fragments) in a single host cell—but not both chains in single host cell: “The evidence of record thus demonstrates that Axel describes nothing more than what is inherently required by the [Cabilly I patent]—production of one desired polypeptide (e.g., a heavy or a light immunoglobulin chain polypeptide) in one transformed host cell.” Ex. 1024, Appeal Brief, at 47.

ii. Owners Argue that the Axel Patent Does Not Disclose the Co-Expression of “One or More” Genes of Interest

Owners also argued that neither Axel nor any other piece of prior art taught or suggested the transformation of a single host cell with any two different genes of interest encoding two different “proteins of interest”—and specifically failed to do so for immunoglobulin heavy and light chains. In Owners’ view, Axel in particular did not disclose the introduction of “more than one desired gene” or “multiple DNA sequences” encoding “different polypeptides of interest” into a

single host cell. Ex. 1024, Appeal Brief, at 49; Ex. 1022, Owners' Resp. (10/30/06), at 44. Such a disclosure of more than one gene in Axel would have been

necessary to support the Office's assertions that the Axel process specifically teaches production of intact antibodies, because only that interpretation leads to the possibility that two different polypeptides (i.e., the heavy and light chains of the immunoglobulin) would be produced by the Axel process.

Ex. 1022, Owners' Resp. (10/30/06), at 44, n.26 (emphasis added). According to Owners, the Axel patent's specific disclosure of a two DNA system (DNAs I and II) did not fill in this alleged gap in the prior art because although DNA I encoded a single protein of interest, DNA II encoded only a "selectable marker" protein and not a second protein "of interest." Ex. 1010, Owners' Resp. (5/21/07), at 21. A POSITA reading the entire disclosure of Axel would therefore "not read the passing references in Axel to 'antibodies' to mean that an antibody tetramer is to be produced by co-expressing the heavy and light chains in one host cell." *Id.*

Owners eventually successfully convinced the PTO that Axel failed to disclose the "co-expression" requirement of the '415 patent claims, (notwithstanding that the host cells in Axel produced two separate proteins). Ex. 1025, NIRC, at 4 ("Axel et al taught a process for inserting foreign DNA into eukaryotic cell by cotransformation with the disclosed foreign DNA I and DNA II

that encodes a selectable marker. Axel et al did not teach a single host cell transformed with immunoglobulin heavy chain and light chain independently. Axel et al did not teach co-expression of two foreign DNA sequences.”). A reexamination certificate issued on May 19, 2009. Ex. 1026, Reexam Cert.

C. Person of Ordinary Skill in the Art

A POSITA at the time of the earliest effective filing date of the '415 patent would have a Ph.D. in molecular biology (or a related discipline, such as biochemistry) with 1 or 2 years of post-doctoral experience, or an equivalent amount of combined education and laboratory experience. The POSITA would also have experience using recombinant DNA techniques to express proteins and familiarity with protein chemistry, immunology, and antibody production, structure, and function. Ex. 1006, Baron Decl. ¶ 28.

D. Claim Construction

The Board is charged with applying the “broadest reasonable interpretation consistent with the specification,” reading the claim language in light of the specification as it would be understood by a POSITA. *In re Cuozzo Speed Techs., LLC*, 793 F.3d 1268, 1275-79 (Fed. Cir. 2015). The terms in the challenged claims of the '415 patent should therefore be given their broadest reasonable interpretation consistent with the specification. Petitioner does not believe that any special meanings apply to the claim terms in the '415 patent. Petitioner's position

regarding the scope of the challenged claims should not be taken as an assertion regarding the appropriate claim scope in other adjudicative forums where a different claim interpretation standard may apply.

IV. RELEVANT PRIOR ART

A. Technology Background

1. The Sophistication of Recombinant DNA Technology Was Advanced by April 8, 1983, and Mammalian Proteins Were Being Made in Host Cells Transformed with Foreign Genes

The technology and associated methodologies for creating, introducing, and expressing (i.e., transcribing and translating) foreign DNA in host cells was past its formative years by April 1983. Ex. 1006, Baron Decl. ¶ 47. The '415 patent notes that by then, “[r]ecombinant DNA technology [had] reached sufficient sophistication that it includes a repertoire of techniques for cloning and expression of gene sequences.” Ex. 1001, 4:7-9. “Various DNA sequences can be recombined with some facility, creating new DNA entities capable of producing heterologous protein product in transformed microbes and cell cultures. The general means and methods . . . for producing expression vectors, and for transforming organisms are now in hand.” *Id.* at 4:9-16. The “expression vector is useful to produce the polypeptide sequence for which the inserted gene codes, a process referred to as ‘expression.’ The resulting product may be obtained by lysis, if necessary, of the host cell and recovery of the product by appropriate purifications from other proteins.” *Id.* at 4:27-32.

Before the priority filing date of the '415 patent, scientists had already produced a few dozen eukaryotic proteins in bacteria. Ex. 1006, Baron Decl. ¶ 49. Timothy Harris, one of Owners' experts who submitted declarations to the PTO during reexamination, authored a 1983 review article compiling all of the higher eukaryotic (including mammalian) proteins expressed in *E. coli* that had been reported to date. Ex. 1027, Harris, at 163-69; Ex. 1028, Harris Decl. ¶ 16. Among the proteins listed are human insulin and fibroblast interferon, human and bovine growth hormone, rat preproinsulin, chicken ovalbumin, and rabbit β -globin.

The Cohen & Boyer patent was one of the foundational platform technologies available before April 1983 that utilized recombinant DNA to make mammalian proteins in bacterial host cells. The Axel patent was similarly a seminal platform technology that advanced the Cohen & Boyer bacterial host cell method by teaching the production of mammalian proteins in eukaryotic (including mammalian) host cells. Ex. 1006, Baron Decl. ¶ 50.

The recombinant production of heterologous proteins in host cells was so well developed by April 1983 that the '415 patent was able to make broad generalizations about the form in which such proteins are produced and how they may be recovered:

[I]t is common for mature heterologous proteins expressed in *E. coli* to be deposited within the cells as insoluble particles which require cell lysis and solubilization in denaturant to permit recovery. On the

other hand, proteins under proper synthesis circumstances, in yeast and bacterial strains, can be secreted into the medium (yeast and gram positive bacteria) or into the periplasmic space (gram negative bacteria) allowing recovery by less drastic procedures. Tissue culture cells as hosts also appear, in general, to permit reasonably facile recovery of heterologous proteins.

Ex. 1001, 12:39-49.

2. The Prior Art Taught Expression of Single Immunoglobulin Chains

Before April 1983, the technology existed to produce either heavy or light immunoglobulin chains in host cells.¹¹ During the '415 patent reexamination, for example, Owners argued that the Axel patent “describes nothing more than what is inherently required by the [Cabilly I patent]—production of one desired polypeptide (e.g., a heavy or a light immunoglobulin chain polypeptide) in one transformed host cell.” Ex. 1024, Appeal Brief, at 47. Similarly, Owners

¹¹ By April 1983, there were a dozen or so published sequences of isolated DNA encoding for at least the variable domains of immunoglobulin heavy and light chains. Ex. 1029, Kabat at 246, 248, 249 (selected pages from a compendium of “sequences of proteins of immunological interest”). A POSITA therefore would have had access to or have been able to isolate without undue experimentation these DNA sequences. Ex. 1006, Baron Decl. ¶ 48 & n.4.

summarized Moore as calling for the “production of heavy and light immunoglobulin polypeptides in separate host cells, and propos[ing] assembly of the multimeric immunoglobulin complex by combining the individually produced chains in a test tube.” Ex. 1023, Owners’ Resp. (6/6/08), at 25; Ex. 1006, Baron Decl. ¶ 52.

Even in 1977, before the Cohen & Boyer patent was filed, Stanley Cohen anticipated that bacteria could be engineered to make antibodies:

[R]ecombinant DNA techniques potentially permit the construction of bacterial strains that can produce biologically important substances such as antibodies and hormones. Although the full expression of higher organism DNA that is necessary to accomplish such production has not yet been achieved in bacteria, the steps that need to be taken to reach this goal are defined, and we can reasonably expect that the introduction of appropriate “start” and “stop” control signals into recombinant DNA molecules will enable the expression of animal cell genes.

Ex. 1030, Cohen, at 655.

In 1979, Arthur Riggs (a co-inventor on the ’415 patent) and Keiichi Itakura wrote that “[c]learly there is no fundamental barrier to prevent transcription and translation [i.e., expression] of eukaryotic genes in prokaryotes.” Ex. 1003, Riggs & Itakura, at 537. “Techniques have developed rapidly, so that the genes necessary for altering the bacteria can be made and inserted with relatively modest

expenditures of time and money.” *Id.* at 533. The authors envisioned that “bacteria may . . . be used for the production of the antibody peptide chains.” *Id.* at 537.

Scientists subsequently employed these methods to produce single immunoglobulin chains in host cells. Rice & Baltimore (Ex. 1020), Ochi (I) (Ex. 1021) and Oi (Ex. 1031) reported experiments in which light chain DNA was successfully transformed into and expressed in mammalian host cells. Ex. 1006, Baron Decl. ¶ 55; Ex. 1016, Office Action (8/16/06), at 5, 23, 26.

The prior art reviewed in the ’415 patent also taught that heavy and light chains produced in separate bacterial host cells may then be assembled *in vitro* using prior art protein denaturing (by reduction) and renaturing (by oxidation) chemical techniques. Ex. 1001, 12:58-13:52; Ex. 1006, Baron Decl. ¶ 56; *see also* Ex. 1003, Riggs & Itakura, at 537-38 (“Bacteria may then be used for the production of the antibody peptide chains, which could be assembled *in vitro*”); Ex. 1019, Moore, 11:1-6 (separately expressed single chains combined *in vitro*); Ex. 1032, Kaplan, 10:31-33 (same).

3. The Prevailing Mindset by April 1983 Was That One or More Proteins of Interest Could be Made in a Single Host Cell

In April of 1983, there was not a “prevailing mindset” in the prior art that only one protein of interest could be made per host cell. There were multiple

references available before April 1983 teaching that more than one mammalian gene could be introduced into and expressed by a single host cell. Ex. 1006, Baron Decl. ¶¶ 57-63 (discussing the prior art summarized below).

For example, U.S. Patent No. 4,487,835 (Ex. 1033) summarizes the state of bacterial expression of eukaryotic (mammalian) proteins before April 1983:

It is known to prepare useful polypeptides and proteins, for example enzymes, hormones . . . by cultivation of bacteria carrying plasmids with genes coding for the desired polypeptides or proteins. It is also known to construct plasmids containing desired genes by so-called recombinant DNA technique, which makes it possible to obtain, from the cultivated bacteria carrying such recombinant DNA plasmids, gene products which inherently are characteristic to other organisms than the bacteria used as host cells. In the preparation of recombinant DNA, a so-called cloning vector, that is, a plasmid which is able to replicate in the host bacterium, is combined with a DNA fragment containing a gene or genes coding for the desired product or products If the foreign DNA is transcribed and translated in the bacterial host, the gene products of the foreign DNA are produced in the bacterial host.

Ex. 1033, 1:17-31, 51-53; *see also* U.S. Patent No. 4,371,614 (Ex. 1034), 1:43-58 (“[O]ne or more genes from a donor organism, such as a . . . eukaryotic cell are introduced into a vector” that is transformed into “a host organism, usually a prokaryotic bacterial microorganism” to “produce corresponding enzymes using

the available protein-synthesizing apparatus of the host.”); U.S. Patent No. 4,762,785 (Ex. 1035), 2:66-3:5 (vector for transforming a prokaryotic host in which “[o]ne or more segments of alien DNA will be included in the plasmid, normally encoding one or more proteins of interest . . . derived from any convenient source, either prokaryotic or eukaryotic, including . . . mammals.”); U.S. Patent No. 4,476,227 (Ex. 1036), 3:1-4 (vector comprising foreign DNA, wherein “the foreign DNA can be of eukaryotic or prokaryotic origin and might include . . . one or more genes for expression and production of commercially useful products”); U.S. Patent No. 4,362,867 (Ex. 1037), 8:48-52 (eukaryotic DNA inserted into a plasmid for transforming *E. coli* to produce a desired protein may include a “gene or genes coding for the cellular production of a desired [protein] product or products”).

Similarly, U.S. Patent No. 4,396,601 (Ex. 1038) teaches introducing and coexpressing multiple independent eukaryotic genes in a single mammalian host cell. The patent teaches that “when two or more genes are to be introduced they may be carried on a single chain, a plurality of chains, or combinations thereof.” Ex. 1038, 3:51-53. “The DNA employed may provide for a single gene, a single set of genes, e.g., the beta-globin gene cluster, or a plurality of unrelated genes.” *Id.* at 5:26-29 (emphasis added). The Southern prior art publication (Ex. 1004), one of Petitioner’s references underlying its grounds for rejection, also teaches

expressing multiple genes of interest in a mammalian host cell by using two vectors to co-transform the cell, with each vector containing a different gene of interest. *Infra* at 32-34, 38-41.

The expression of “one or more genes,” “two or more genes,” a “plurality of unrelated genes” or “a gene or genes” “encoding one or more proteins of interest” in a single host cell—this was the prevailing mindset in April 1983, and not the “one polypeptide per host cell” postulate advocated by Owners during reexamination. Ex. 1006, Baron Decl. ¶ 64. And it was this prevailing mindset that is reflected around the time of filing of the ’415 patent in the teachings of heavy and light chain co-expression in the Boss patent (Ex. 1012, 5:43-56, 6:1-17) and in the work of scientists who published their heavy and light chain co-expression experiments shortly after April 1983 (Ex. 1040, Ochi (II)). Ex. 1006, Baron Decl. ¶ 64.

The state of the art of the co-expression of genes (eukaryotic and otherwise) in recombinant systems before April 1983 was advanced enough so that even as early as 1980, Dr. César Milstein—a Nobel Laureate (with Georges Köhler) for his work on monoclonal antibodies—suggested its application in antibody production. He anticipated bacterial and mammalian host cells transformed with heavy and light chain DNA, followed by expression of the respective polypeptides. Ex. 1039, Milstein, at 409-10. Dr. Milstein observed that if bacterial host cells are used, “we

have to face the possibility that bacteria may not be able to handle properly the separated heavy and light chains so that correct assembly becomes possible.” *Id.* at 410. This concern of Dr. Milstein—that bacteria may not be able to correctly assemble the heavy and light chains—necessarily presumes a single bacterial cell that has been transformed with the both heavy chain and light chain genes and was co-expressing both genes. Ex. 1006, Baron Decl. ¶ 65.

Thus, in the five years preceding the ’415 patent’s filing date: (1) there was an available set of platform technologies for making mammalian proteins in bacterial and mammalian host cells; (2) the ability to make single immunoglobulin (either heavy or light) chains in bacterial and mammalian host cells was known in the art; and (3) the art expressed multiple suggestions that more than one gene of interest can be introduced into a host cell to produce more than one protein of interest. All of these teachings are germane to Petitioner’s grounds for rejection of the challenged claims.

B. References Underlying the Grounds for Rejection

1. Bujard Teaches Introducing and Expressing a “Plurality of Genes” in Bacterial or Mammalian Host Cells and Identifies “Immunoglobulins” as a Protein of Interest

Bujard (Ex. 1002) issued on January 22, 1985, to inventors Hermann Bujard and Stanley Cohen based on an application filed May 20, 1981. Bujard qualifies as prior art under § 102(e). Bujard was never cited by Owners, or identified or relied

upon by the PTO, during prosecution or reexamination of the '415 patent Bujard is directed to vectors made by recombinant DNA technology for expressing proteins of interest in transformed host cells. Ex. 1006, Baron Decl. ¶¶ 66-69. Bujard notes that the preexisting technology had already “established the feasibility of producing a wide variety of naturally occurring and synthetic polypeptides by means of hybrid DNA technology,” but acknowledges that “there are continuing and extensive efforts to provide for more efficient and economic methods for producing the polypeptides.” Ex. 1002, 1:13-18. The vectors in Bujard are optimized over prior art vectors by increasing their efficiency in transcribing DNA to RNA and in expressing one or more genes of interest (referred to in the patent as “structural genes”¹²) in host cells to produce one or more proteins of interest. Ex. 1006, Baron Decl. ¶¶ 66-67, 69-71. Bujard identifies immunoglobulins among the proteins that can be made by the disclosed process, vectors, and transformed host cells taught by Bujard. Ex. 1006, Baron Decl. ¶ 74.

The vector of Bujard consists of a “strong promoter, followed by a DNA sequence of interest, optionally followed by one or more translational stop codons in one or more reading frames, followed by a balanced terminator, followed by a

¹² A “structural gene” is a gene that “provid[es] a poly(amino acid),” i.e., a protein. Ex. 1002, 3:9-14; Ex. 1060, Baron Decl. ¶ 70.

marker allowing for selection of transformants.” Ex. 1002, 2:3-20; Ex. 1006, Baron Decl. ¶ 68. The DNA sequence of interest, which “usually” consists of “structural genes,” is inserted between the strong promoter and terminator to “provide for efficient transcription and/or expression of the sequence.” Ex. 1002, 2:33-38; Ex. 1006, Baron Decl. ¶¶ 68-71. The DNA sequence of interest may contain “more than one gene, that is, a plurality of genes, including multimers and operons.”¹³ Ex. 1002, 3:46-48; Ex. 1006, Baron Decl. ¶¶ 69-71; *see also* Ex. 1002, 7:61-63 (“[O]ne or more structural genes may be introduced between the promoter and terminator”) and 8:7-11 (“Alternatively, the gene(s) of interest may be ligated to the appropriate regulatory signal sequences before insertion into the [plasmid] vehicle”). These are the exact teachings of multiple DNA sequences that Owners

¹³ “Multimer” refers to a protein with more than one subunit. *Supra* at 13, n.8. In the context of Bujard, a POSITA would therefore understand the use of the term “multimer” to mean “genes encoding multimeric proteins.” Ex. 1060, Baron Decl. ¶¶ 72-73. When a multimeric protein is encoded by a “plurality of genes,” with each gene making a different type of polypeptide, this can only be construed as a multimeric protein with chemically distinct (i.e., non-identical) polypeptide subunits, for example, an immunoglobulin. Ex. 1060, Baron Decl. ¶¶ 73-74.

argued during the '415 patent reexamination were absent from the Axel patent.
Supra at 16-17.

Further, the vector with the inserted DNA sequence of interest containing one or more structural genes “can be used with one or more hosts for gene expression” of a “wide variety of poly(amino acids)” by transforming the host cell (either a microorganism, e.g., *E. coli*, or a mammalian cell) with the vector. Ex. 1002, 3:61-63, 6:23-37, 8:1-3, 11:28-31; Ex. 1006, Baron Decl. ¶ 69. Among the “wide variety” of genes and proteins of interest identified in the patent are “immunoglobulins e.g. IgA, IgD, IgE, IgG and IgM and fragments thereof,” as well as “free light chains.” Ex. 1002, 4:14-16, 4:30-36, 5:11-27. The Bujard patent makes clear the common knowledge at the time that antibodies are assembled from multiple, discrete polypeptides (four—two heavy and two light chains) encoded for by two different genes: it identifies the molecular formula of each type of immunoglobulin that can be produced according to the disclosed method—for example, the patent notes that IgG has the molecular formula of “ $\gamma_2\kappa_2$ or $\gamma_2\lambda_2$ ” (two heavy chains (γ_2) and two light chains (κ_2 or λ_2)). Ex. 1002, 5:11-14; Ex. 1006, Baron Decl. ¶ 75.

The resultant proteins produced by the transformed host cells may be prepared either “as a single unit” or “as individual subunits and then joined together in appropriate ways.” Ex. 1002, 4:19-21. The “single unit” is a reference

that a POSITA would understand to include an *in vivo* assembled multimeric protein, such as an immunoglobulin; the joining together of “individual subunits” by “appropriate ways” is a reference that a POSITA would understand to include the *in vitro* assembly of the constituent polypeptide subunit chains of a multimeric protein, such as an immunoglobulin. Ex. 1006, Baron Decl. ¶ 78.

2. Riggs & Itakura Teaches Hybridomas as a Source of Antibody Genes and the *In Vitro* Assembly of Heavy and Light Chains

Riggs & Itakura (Ex. 1003) published in 1979 and qualifies as prior art under §102(b). Riggs & Itakura was never cited by Owners, or identified or relied upon by the PTO, during prosecution or reexamination of the '415 patent.

Arthur Riggs and Keiichi Itakura were among the first scientists to use recombinant DNA technology to express mammalian proteins in bacteria. Ex. 1006, Baron Decl. ¶ 90. In the article, they provide an overview of their work on making human insulin in bacteria: creating synthetic DNA encoding for the insulin A and B polypeptide chains, using recombinant DNA techniques to insert the genes into separate plasmids, separately transforming *E. coli* cells with plasmids containing the genes for the A and B chains, recovering the expressed chains from lysed bacterial cells, and *in vitro* assembly of the chains into an intact insulin molecule. Ex. 1003, at 531-33; Ex. 1006, Baron Decl. ¶ 90. The authors saw the practical application for this technology as extending beyond insulin

production. Ex. 1003, at 537-38; Ex. 1006, Baron Decl. ¶ 90. They taught that “[h]ybridomas will provide a source of mRNA for specific antibodies. Bacteria may then be used for the production of the antibody peptide chains, which could be assembled *in vitro* and used for passive immunization.” Ex. 1003, at 537-38; Ex. 1006, Baron Decl. ¶ 90.

3. Southern Teaches One Host Cell Transformed with Two Vectors

Southern (Ex. 1004) published in July 1982 and qualifies as prior art under §102(a). Southern was never cited by Owners, or identified or relied upon by the PTO, during prosecution or reexamination of the ’415 patent.

Southern teaches a single mammalian host cell that is “cotransformed”¹⁴ with two separate plasmids: the first (called “pSV2neo”) containing the selectable marker gene “neo,” which when expressed as a protein provides the cells with the ability to grow in the presence of the antibiotic G418; the second (“pSV2gpt”) containing the selectable marker gene “gpt,” which when expressed as a protein provides the cells with the ability to grow in the presence of the antibiotic MPA. Ex. 1004, at 336-37, Table 3; Ex. 1006, Baron Decl. ¶¶ 91-92. The cotransformed host cells successfully expressed both selectable marker proteins and were able to

¹⁴ This is also referred to as “cotransduction” and “cotransfection” in the article. Ex. 1004, at 336-37; Ex. 1060, Baron Decl. ¶ 93, n.7.

grow in the presence of both antibiotics, i.e., they were “double selected.” Ex. 1004, at 336-37, Table 3; Ex. 1006, Baron Decl. ¶¶ 93-94. Southern evinces a formal proof that the two expression vectors are compatible and can be used and selected for simultaneously in the same cell without interfering with each other. Ex. 1006, Baron Decl. ¶ 93.

While Southern’s cotransformation experiments used the two vectors without “gene-of-interest” insertions, that was merely an experimental convenience. Ex. 1006, Baron Decl. ¶ 94. Both vectors are described repeatedly as expression vectors, and the intent to use them to coexpress multiple “genes of interest,”¹⁵ one on each vector, with double selection is made explicit in the paper’s concluding statement: “[c]otransformation with nonselectable genes can be accomplished by inserting genes of interest into vector DNAs designed to express neo or gpt. The schemes used to select for the expression of gpt and neo are complementary and experiments that exploit the possibilities of a double and dominant selection are now in progress.” Ex. 1004, at 339; Ex. 1006, Baron Decl. ¶ 94.

¹⁵ Also called “nonselectable genes” in Southern, that is, genes that do not confer a selective advantage to the host cell. Ex. 1004, at 336; Ex. 1060, Baron Decl. ¶ 95, n.8.

V. FULL STATEMENT OF PRECISE RELIEF REQUESTED AND THE REASONS THEREFOR (37 C.F.R. § 42.22(a))

A. Explanation of Ground 1 for Unpatentability: Claims 1, 3, 4, 11, 12, 14, 19, and 33 Are Obvious Over Bujard in View of Riggs & Itakura

Claims 1, 3, 4, 11, 12, 14, 19, and 33 are invalid as obvious over Bujard in view of Riggs & Itakura. Independent claims 1 and 33 require the recombinant production of an immunoglobulin molecule (i.e., an antibody) or immunologically functional fragment by “independently expressing” DNA sequences encoding at least the variable domains of the immunoglobulin heavy and light chains within a “single host cell.” Dependent claim 3 requires that the heavy and light chain DNA sequences be “present in a single vector” and dependent claim 4 requires that the “vector” of the claims from which it depends be a “plasmid.” Dependent claim 11 requires that the DNA sequences encode the “complete” heavy and light chain polypeptides and dependent claim 12 requires that any “constant domain” encoded by the DNA sequences be “derived from the same source as the variable domain to which it is attached.” Dependent claim 14 also requires that the heavy and light chain DNA sequences be “derived from one or more monoclonal antibody producing hybridomas” and dependent claim 19 requires that the host cell of claim 1 is a “mammalian cell.”

The Bujard patent teaches a process for producing proteins of interest—among which the patent expressly identifies immunoglobulins—in a transformed

host cell using a plasmid vector that is optimized to increase the efficiency of expression. Ex. 1002, 2:1-20, 3:9-14, 3:61-62, 4:14-16, 4:30-36, 5:11-27; Ex. 1006, Baron Decl. ¶ 96; *see also supra* at 27-31. Producing such proteins as taught by Bujard occurs in a single host cell—either bacterial or mammalian—that is transformed with a single plasmid containing “more than one gene, that is, a plurality of genes.” Ex. 1002, 3:46-48, 3:61-62, 6:23-37; Ex. 1006, Baron Decl. ¶ 96; *see also supra* at 28-30.

A POSITA would have been motivated to combine Bujard with the *in vitro* assembly disclosures in Riggs & Itakura, (Ex. 1003, at 537-38), with a reasonable expectation of success in achieving the purported invention of the challenged claims, thus rendering the claims obvious. Ex. 1006, Baron Decl. ¶ 104.

Bujard and Riggs & Itakura are publications in the same general field of research: the production of heterologous eukaryotic proteins in host cells. Ex. 1006, Baron Decl. ¶ 105. Beyond that general motivation to combine the references, the particular motivation to combine the *in vitro* protein assembly techniques of Riggs & Itakura with the Bujard antibody production method is found in Bujard itself. Ex. 1006, Baron Decl. ¶ 105. Bujard suggests at least two ways of obtaining the desired protein end product, one of which is that “individual [protein] subunits” can be “joined together in appropriate ways.” Ex. 1002, 4:20-21; Ex. 1006, Baron Decl. ¶ 105; *see also supra* at 31. When the desired end

product is a multi-subunit protein such as an immunoglobulin, a POSITA would have understood that the individual subunits (heavy and light chains) may be recombined according to known methods, including those referenced in Riggs & Itakura, which addresses the same problem of joining unassociated immunoglobulin (and insulin) chains separately produced in microorganism host cells. Ex. 1003, at 537-38; Ex. 1006, Baron Decl. ¶ 105. A POSITA would therefore have had a good reason to combine Bujard with Riggs & Itakura. Ex. 1006, Baron Decl. ¶ 105.

A POSITA would have also reasonably predicted that combining Bujard with the *in vitro* assembly techniques in Riggs & Itakura would result in an assembled immunoglobulin molecule. Ex. 1006, Baron Decl. ¶ 106. Riggs & Itakura themselves demonstrated that the separate chains of insulin could be joined *in vitro*, and taught that the same or similar techniques could be used successfully for immunoglobulin chains made by recombinant DNA means in microorganism host cells. Ex. 1003, at 531-32, 537-38; Ex. 1006, Baron Decl. ¶ 106. There would have been no reason for a POSITA to believe that these methods could not also be successfully used to assemble the heavy and light chains produced by Bujard's similar recombinant DNA methodologies. Ex. 1006, Baron Decl. ¶ 106. A POSITA would therefore have had a reasonable expectation of success in combining Bujard with Riggs & Itakura to result in the subject matter of the

challenged claims. Ex. 1006, Baron Decl. ¶ 106. In sum, a POSITA would have found it obvious to insert the genes encoding for the heavy and light chains, separated by a stop codon, between the promoter and terminator sequences of the vector, which would permit the independent expression of those genes as separate molecules in the transformed host cell. *See* Ex. 1006, Baron Decl. ¶ 97.

As noted above, dependent claim 14 requires that the heavy and light chain DNA sequences be “derived from one or more monoclonal antibody producing hybridomas.” A POSITA would have been motivated to combine Bujard with the hybridoma teachings in Riggs & Itakura (Ex. 1003, at 537) with a reasonable expectation of success in achieving the purported invention of claim 14, thus rendering the claim obvious. Ex. 1006, Baron Decl. ¶ 107. Riggs & Itakura expressly teaches that hybridomas would be a source of genetic material for heavy and light chains, which could then be used for their production in bacteria. Ex. 1003, at 537; Ex. 1006, Baron Decl. ¶ 107. As discussed above, a POSITA would have been motivated to combine these references, and would have done so with the reasonable expectation that the hybridoma immunoglobulin genes could be successfully used in the Bujard system to result in the subject matter of claim 14. Ex. 1006, Baron Decl. ¶ 107. Indeed, a POSITA would not doubt that immunoglobulin genes derived from a hybridoma would work in the Bujard method, and Riggs & Itakura itself teaches that bacterial host cells could be used to

successfully make the chains from these genes. Ex. 1003, at 537-38; Ex. 1006, Baron Decl. ¶ 107.

B. Explanation of Ground 2 for Unpatentability: Claims 1, 2, 18, 20, and 33 Are Obvious Over Bujard in View of Southern

Claims 1, 2, 18, 20, and 33 are invalid as obvious over Bujard in view of Southern. Claim 2 requires that the two DNA sequences of claim 1 “are present in different vectors”; claims 18 and 20 require a host cell and mammalian host cell, respectively, transformed with both of these separate vectors. A POSITA would have been motivated to combine (1) Bujard’s teaching of a mammalian host cell transformed with two DNA sequences (for heavy and light chains), both in a single vector with (2) the co-transformation approach taught in Southern, i.e., a mammalian host cell transformed with two vectors, each with a different selectable marker and gene of interest. Ex. 1006, Baron Decl. ¶ 108. Both Bujard and Southern are publications directed to the expression of heterologous proteins in cells by using recombinant DNA technology and the related tools (vectors, host cells) to accomplish this. Ex. 1006, Baron Decl. ¶ 108. Beyond this general motivation to combine the references, a POSITA would have recognized that both references have as a goal the expression of genes of interest in a single transformed host cell, whether by using one (Bujard) or two (Southern) vectors. Ex. 1006, Baron Decl. ¶ 108. A POSITA would therefore have had a reason to combine Bujard with Southern and to modify Bujard accordingly by splitting the heavy and

light chain DNA sequences into two separate vectors to be transformed in a single mammalian host cell. Ex. 1006, Baron Decl. ¶ 108.

A POSITA would have also reasonably predicted that this modification of Bujard in accordance with Southern would have resulted in the purported inventions of claims 2, 18, and 20. Ex. 1006, Baron Decl. ¶ 109. A POSITA would have been confident that a host cell's expression (transcription and translation) machinery would successfully make heavy and light chains from DNA sequences in separate vectors based on Southern's teaching that multiple proteins (selectable markers and proteins of interest) present on separate vectors could be expressed in a single host cell. Ex. 1006, Baron Decl. ¶ 109. Once a POSITA knows that heavy and light chain genes could be successfully co-expressed in a single host cell when present on one vector (as taught by Bujard), and that two genes of interest could also be successfully expressed in a single host cell when present on two vectors (as taught by Southern), the POSITA would have been confident that heavy and light chains could be successfully co-expressed in a single host cell when present on separate vectors. Ex. 1006, Baron Decl. ¶ 109. A POSITA would have known that the expression machinery in cells works universally, regardless of any difference in genes (heavy/light chain versus non-immunoglobulin polypeptides) or whether they are on separate vectors (instead of one). Ex. 1006, Baron Decl. ¶ 109. Furthermore, because the heavy and light

chain genes are on different vectors in the same host cell, they would necessarily be “independently expressed” and produced as “separate molecules,” as required by claim 2. Ex. 1006, Baron Decl. ¶ 109. A POSITA would therefore have had a reasonable expectation of success in combining Bujard with Southern to result in the subject matter of challenged claims 2, 18, and 20. Ex. 1006, Baron Decl. ¶ 109.

Because claim 2 is obvious over Bujard in combination with Southern, independent claim 1 on which claim 2 depends is necessarily obvious as well. *Callaway Golf Co. v. Acushnet Co.*, 576 F.3d 1331, 1344 (Fed. Cir. 2009) (“A broader independent claim cannot be nonobvious where a dependent claim stemming from that independent claim is invalid for obviousness.”); Ex. 1006, Baron Decl. ¶ 110. Furthermore, because the scope of claim 33 is no different in any meaningful way than the scope of claim 1—i.e., they are both directed to coexpression of heavy and light chains in a single host cell, and are broad enough to encompass this through either a single vector or two vector transformation—claim 33 is similarly obvious (as explained above for claim 2) over Bujard in view of Southern. Ex. 1006, Baron Decl. ¶ 110.

C. Secondary Indicia of Non-Obviousness in the Public Record Do Not Rebut Petitioner’s Prima Facie Case of Obviousness

During the reexamination of the ’415 patent, Owners relied upon the “licensing record and commercial success” of the patent, asserting that it provided

evidence of non-obviousness of the claims. Ex. 1023, Owners' Resp. (6/6/08), at 40-42; Ex. 1046, Walton Decl., at 4-9. Neither Owners nor Dr. Walton provided any "explanation or evidence to establish [a] nexus between the merits of the invention and the licenses themselves" or to the licensing royalties received by Owners. See *CBS Interactive Inc. v. Helferich Patent Licensing, LLC*, IPR2013-00033, Paper 21, at 22 (Mar. 25, 2013) (citing *Iron Grip Barbell Co. v. USA Sports, Inc.*, 392 F.3d 1317, 1324 (Fed. Cir. 2004); *SIBIA Neurosciences, Inc. v. Cadus Pharm. Corp.*, 225 F.3d 1349, 1358 (Fed. Cir. 2000)). There is no explanation of "the terms of the licenses and the circumstances under which they were granted," for example, whether "they were entered into as business decisions to avoid litigation or other economic reasons." See *CBS Interactive*, IPR2013-00033, Paper 21, at 22. For these reasons alone, any reliance here by Owners on evidence from the reexamination of alleged licensing acquiescence and commercial success should be given no weight. See *id.*

Moreover, the history of licensing and licensing revenues relied on in the reexamination is now stale (there is no information beyond 2007 in Dr. Walton's declaration) and does not reflect the pharmaceutical and biotech industry's recent collective opinion of the value of the '415 patent or its validity. Since the '415 patent issued, Owners have been involved in six patent infringement lawsuits challenging the validity of the '415 patent, only one of which was filed before

2007, and all of which settled after 2007.¹⁶ Many of these challenges covered antibody licenses that Owners highlighted during the reexamination proceeding as evidencing acquiescence by the industry. Ex. 1046, Walton Decl., at ¶ 29, n.6; Ex. 1041, Walton Expert Rep., at 23 (exhibit to Request for Reconsideration (Ex. 1042)). Owners' assurances of the industry's acquiescence to the '415 patent before 2007 cannot be squared with the subsequent challenges brought by the very types of "large, sophisticated, patent-savvy companies," (Ex. 1023, Owners' Resp. (6/6/08), at 41), who Owners claim demonstrate their respect for the '415 patent. Owners' decisions to settle with each of these challengers before a court could render a decision on the invalidity arguments presented paints a different picture than the speculation put forth by Dr. Walton as to why these companies entered into licenses in the first place.

¹⁶ See complaints and dismissals for the '415 patent in lawsuits involving MedImmune (Exs. 1047, 1048), Centocor (Exs. 1049, 1050), Glaxo Group Ltd. (Exs. 1051, 1052), Human Genome Sciences (Exs. 1053, 1054), Eli Lilly (Exs. 1055, 1056), and Bristol-Myers Squibb (Exs. 1057, 1058) filed against Genentech and City of Hope.

VI. MANDATORY NOTICES UNDER 37 C.F.R. § 42.8(A)(1)

A. Real Party-In-Interest Under 37 C.F.R. § 42.8(b)(1)

Pursuant to 37 C.F.R. § 42.8(b)(1), Petitioner certifies that Merck Sharp & Dohme Corp. is the real party-in-interest.

B. Related Matters Under 37 C.F.R. § 42.8(b)(2)

Pursuant to 37 C.F.R. § 42.8(b)(2), Petitioner states that five IPR petitions for the '415 patent have been filed: IPR2015-01624, IPR2016-00383, IPR2016-00460, IPR2016-00710, and IPR2016-01373. IPR2015-01624 was instituted by the Board on February 5, 2016 and was terminated on September 2, 2016. IPR2016-00460 was instituted and joined with IPR2015-01624 by the Board on June 8, 2016 and was terminated on September 2, 2016. IPR2016-00383 was denied institution by the Board on June 23, 2016. IPR2016-00710 was filed on March 3, 2016 instituted on September 8, 2016. IPR2016-01373, which was filed by Petitioner, was filed July 7, 2016, and the Board has not issued an institution decision in that proceeding.

C. Lead and Back-up Counsel and Service Information Under 37 C.F.R. § 42.8(b)(3), (4)

Petitioner hereby designates lead and backup counsel as follows:

Lead Counsel	Backup Counsel
Raymond N. Nimrod (Reg. No. 31,987) QUINN EMANUEL URQUHART & SULLIVAN, LLP 51 Madison Ave., 22 nd Floor	Matthew A. Traupman (Reg. No. 50,832) QUINN EMANUEL URQUHART & SULLIVAN, LLP 51 Madison Ave., 22 nd Floor

New York, NY 10010 General Tel: (212) 849-7000 Direct Tel: (212) 849-7412 Fax: (212) 849-7100 raynimrod@quinnemanuel.com	New York, NY 10010 General Tel: (212) 849-7000 Direct Tel: (212) 849-7322 Fax: (212) 849-7100 matthewtraupman@quinnemanuel.com Katherine A. Helm (<i>pro hac vice</i> to be filed) SIMPSON THACHER & BARTLETT LLP 425 Lexington Avenue New York, NY 10017 General Tel: (212) 455-2000 Direct Tel: (212) 455-3647 Facsimile: (212) 455-2502 khelm@stblaw.com
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Pursuant to 37 C.F.R. § 42.10(b), a Power of Attorney has been filed herewith. Petitioner consents to electronic service at the email addresses above. Petitioners will request authorization to file a motion for Katherine A. Helm to appear *pro hac vice*. Dr. Helm is an experienced attorney and has an established familiarity with the subject matter at issue in this proceeding. Petitioners intend to file a motion seeking the admission of Katherine A. Helm to appear *pro hac vice* when authorized to do so.

VII. PAYMENT OF FEES (37 C.F.R. § 42.15(A) AND § 42.103))

The required fees are submitted herewith in accordance with 37 C.F.R. §§ 42.103(a) and 42.15(a). If any additional fees are due during this proceeding, the Office is authorized to charge such fees to Deposit Account No. 505708. Any overpayment or refund of fees may also be deposited in this Deposit Account.

VIII. CONCLUSION

Petitioner submits that issues have been presented that demonstrate a reasonable likelihood that claims 1-4, 11-12, 14, 18-20, and 33 of the '415 patent are unpatentable in view of the prior art. Petitioner therefore requests that the Board institute *inter partes* review for each of those claims.

DATED: October 11, 2016

Respectfully submitted,

By s/ Matthew A. Traupman
Raymond N. Nimrod (Reg. No. 31,987)
raynimrod@quinnemanuel.com
Matthew A. Traupman (Reg. No. 50,832)
matthewtraupman@quinnemanuel.com
QUINN EMANUEL URQUHART
& SULLIVAN LLP
51 Madison Avenue, 22nd Floor
New York, NY 10010
Tel: (212) 849-7000
Fax: (212) 849-7100

Katherine A. Helm (*pro hac vice* to be filed)
khelm@stblaw.com
SIMPSON THACHER & BARTLETT LLP
425 Lexington Avenue
New York, NY 10017
Telephone: (212) 455-2000
Facsimile: (212) 455-2502

*Attorneys for Petitioner Merck Sharp &
Dohme Corp.*

CERTIFICATE OF COMPLIANCE

I hereby certify that the foregoing Petition for *Inter Partes* Review of U.S. Patent No. 6,331,415 contains 10,469 words as measured by the word processing software used to prepare the document, in compliance with 37 C.F.R. § 42.24(d).

DATED: October 11, 2016

Respectfully submitted,

By s/ Matthew A. Traupman
Matthew A. Traupman (Reg. No. 50,832)
matthewtraupman@quinnemanuel.com
QUINN EMANUEL URQUHART
& SULLIVAN LLP
51 Madison Avenue, 22nd Floor
New York, NY 10010
Tel: (212) 849-7000
Fax: (212) 849-7100

CERTIFICATE OF SERVICE

I hereby certify that true and correct copies of the foregoing Petition for *Inter Partes* Review of U.S. Patent No. 6,331,415 and Exhibits 1001-1058 were served on October 11, 2016 via FEDERAL EXPRESS to the attorneys of record for U.S. Patent No. 6,331,415 as evidenced on Public PAIR on October 11, 2016, namely:

JEFFREY P. KUSHAN, ESQ.
Sidley Austin LLP
1501 K Street, N.W.
Washington, D.C. 20005

and

SEAN JOHNSTON, ESQ.
Genentech, Inc.
1 DNA Way
South San Francisco, CA 94080

DATED: October 11, 2016

Respectfully submitted,

By s/ Matthew A. Traupman
Matthew A. Traupman (Reg. No. 50,832)
matthewtraupman@quinnemanuel.com
QUINN EMANUEL URQUHART
& SULLIVAN LLP
51 Madison Avenue, 22nd Floor
New York, NY 10010
Tel: (212) 849-7000
Fax: (212) 849-7100